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# **Application of Probit Analysis in Studying Allelopathy Phenomenon**

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#### ABSTRACT

Probit analysis is a type of regression used to analyze the relationship between a stimulus and the quantal response. Allelopathy refers to direct or indirect negative or positive effects of one plant on another through the release of chemical compounds into the environment. This study was carried out to apply probit analysis in investigating the allelopathic effects of the leaves aqueous extracts of apple of Sodom [Calotropis procera (Aiton) W.T.] on the inhibition of seed germination of African rattlebox (Crotalaria saltiana Andr.). Laboratory experiments were carried out at the Faculty of Agricultural Sciences, University of Gezira, Sudan, in season 2014/15. Ten concentrations (2.3, 4.6, 7.0, 9.3, 11.6, 13.9, 16.2, 18.5, 20.8 and 23.2 g/l) of leaves aqueous extract of apple of Sodom were prepared from a stock solution (50 g / 1). A control with sterilized-distilled water was included for comparison. Treatments were arranged in completely randomized design with four replicates. The seeds were examined for inhibition in germination at three days after initial germination. Collected data were transformed using Abbott's formula and subjected to probit analysis procedure ( $P \le 0.5$ ). The results showed that the leaves aqueous extract of apple of Sadom had allelopathic on seed germination of African rattlebox with a direct positive relationship between concentration (g/l) and inhibition (%). Also, the data indicated that the plotting of corrected inhibition (%) against concentration (%) formed a sigmoid curve. Probit analysis transformed the sigmoid concentration-response curve to a straight line. Hence, the LC<sub>25</sub> (2.16 g/l),  $LC_{50}$  (8.55 g/l) and  $LC_{75}$  (33.88 g/l) were accurately estimated. It is concluded that probit analysis is an appropriate procedure to study the allelopathy phenomenon.

**Keywords:** African rattlebox, Allelopathy, Apple of Sodom, *Calotropis, Crotalaria*, Probit

تطبيق تحليل البروبيت في دراسة ظاهرة التضاد الحيوي عوض الله بلال دفع الله<sup>1</sup>\* ، هناء عباس حسين<sup>1</sup> ، عبد الرحيم بشير حامد<sup>2</sup> <sup>1</sup> كلية العلوم الزراعية، جامعة الجزيرة، واد مدني، السودان.

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الخلاصة

تحليل الاحتمالية "Probit" هو نوع من الانحدار يستخدم لتحليل العلاقة بين التحفيز والاستجابة الكمية. يشير التضاد الحيوي "Allelopathy" إلى التأثيرات السلبية أو الإيجابية المباشرة أو غير المباشرة لنبات على آخر من خلال إطلاق المركبات الكيميائية في البيئة. أجريت هذه الدراسة من أجل تطبيق تحليل الاحتمالية في دراسة تأثير التضاد الحيوي للمستخلص المائي لأوراق نبات العشر [W.T (Aiton) (Aiton)

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#### Introduction

Probit analysis is a type of regression used to analyze the relationship between a stimulus (concentration) and the quantal (all or nothing) response. The idea of probit analysis was originally proposed since 1934 by Bliss [1]. He offered the idea of transforming the sigmoid concentrationresponse curve to a straight line. The statistical theory and techniques using probit analysis for analyzing data from concentration-quantal response experiments were further developed and discussed in details by Simon [2]. Probit analysis is widely used to analyze data obtained from the bioassays that are generally in percent response at the corresponding concentrations. The response is always binary by nature (ves/no) and the relationship between the response and the concentration is always sigmoid. Probit analysis acts as a transformation from sigmoid to linear and then runs a regression on the relationship. Once a regression is run, the output of the probit analysis could be used to compare the amount of chemical required to create the same response in each of the various chemicals. There are many endpoints used to compare the differences of concentration response. However, LC<sub>50</sub> value is the most widely used outcome of the concentration-response experiments. LC50 is the lethal concentration that is required to kill 50 % of the population. There are several computer programs such as SPSS, SAS, Minitab, etc. that are faster and more accurate in estimating the critical LC<sub>50</sub> value.

Allelopathy refers to direct or indirect negative or positive effects of one plant on another through the release of chemical compounds into the environment [3]. These chemical compounds, known as allelochemicals, are released from plant parts by exudation from roots, leaching and volatilization from stems and leaves, or decomposition of plant material in both natural and agricultural systems [4, 5]. The allelochemicals can reduce cell division or the levels of auxin that induces the growth of shoot and roots [6]. Allelochemicals such as phenolic compounds inhibit root and shoot length [7]. Growth inhibition caused by these allelochemicals may probably be due to its interference with the plant growth processes [6]. Allelochemicals released to the environment can either inhibit shoot and/or root growth, nutrient uptake, or may attack a naturally occurring symbiotic relationship, thereby destroying the plant's source of a nutrient. Understanding well the mechanism of allelopathic interactions between weeds and crops will enable to come up with proper and effective management ways to prevent further infestations. Several statistical analysis procedures were applied to analyze data in studying allelopathy phenomenon, such as descriptive analysis, analysis of variance, correlation and regression.

Apple of Sodom [*Calotropis procera* (Aiton) W.T.], a member of family Asclepiadaceae, is a xerophytic perennial shrub. It is native to tropical and subtropical Africa, Asia and common in the Middle East. It grows on a variety of soils, from fine to coarse texture, with varying degrees of salinity [8]. The widespread and persistent occurrence of apple of Sodom near the agricultural fields causes some adverse effect on the cultivated crops through allelopathic interaction [9]. Thus, the plant has received much attention from researchers due to its allelopathic behavior and has extensively been used for the control of many plants [10]. African rattlebox (*Crotalaria saltiana* Andr.), belonging to the family Fabaceae, is a large and diverse group of the sub-family papilionoidae (legumes) that

largely occur in Africa [11]. This plant is sometimes useful for nitrogen fixation by its root nodules [12]. The plant is common throughout central and northern Sudan [13, 14].

Despite that the idea of probit analysis procedure was proposed and developed since 1934 and the term allelopathy was first used in 1937, however, there is no application of probit analysis in studying the allelopathy phenomenon. Therefore, this study was carried out to apply probit analysis in investigating the allelopathic effects of the leaves aqueous extracts of apple of Sodom on the inhibition of seed germination of African rattlebox.

# **Materials and Methods**

### 1. Experimental site

A series of germination tests were conducted in the Biology laboratory at the Faculty of Agricultural Sciences (FAS), University of Gezira (UofG), Sudan in June, 2018. The laboratory has an average temperature range between 26 - 28°C and the relative humidity ranges between 65 and 68 %.

# 2. Materials collection

Leaves of mature plants of apple of Sodom were collected from the Experimental Farm of the FAS in March, 2018. The leaves were transferred to the Biology laboratory of the FAS. Then, the leaves were washed with sterilized distill water and air dried on bench for 21 days at room temperature and in a dark room to avoid the direct sun light that might cause undesired reactions. The dried leaves were then crushed into powder and kept in brown bottles till used. The seeds of African rattlebox (*Crotalaria saltiana* Andr.) that have a germination percentage of 95-100% were collected from the Experimental Farm of the FAS, Gezira state, Sudan, in season 2017/18. The seeds were surface sterilized by 1% (v/v) solution of sodium hypochlorite (NaOCl), with continuous agitating for 3 min to reduce fungal infections. Subsequently, the seeds were washed with sterilized distill water for several times and stored at room temperature till used.

# 3. Preparation and calculation of the actual concentration of the leaves aqueous extract

Fifty grams, initial weight (IW), of leaves powder of apple of Sodom were placed in a conical flask, sterilized distill water was added to give a volume of 1000 ml, and then the flasks were shaken for 24 hours at room temperature  $(26\pm2^{\circ}C)$  by an orbital shaker (160 rpm). The aqueous extract of the leaves was filtered by a muslin cloth, the leachate was dried, and the final weight (FW) of the filter or the weight of the precipitation (cake) was calculated by a sensitive balance. The final volume (FV) of the water extract for the apple of Sodom leaves was calculated by a measuring cylinder. The actual concentration (AC) of the aqueous extract of the leaves was calculated using the following equation:

$$AC\left(g/l\right) = \frac{(IW - FW) \times 1000}{FW} \tag{1}$$

# 4. Bioassay experiment procedure

Ten concentrations (n) of the leaves aqueous extract were prepared by sequential dilution of the stock extract with sterilized-distilled water to give 2.3, 4.6, 7.0, 9.3, 11.6, 13.9, 16.2, 18.5, 20.8 and 23.2 g/l. A control with sterilized-distilled water was included for comparison. Seeds (100 seeds) of African rattlebox were placed on a Glass Fiber Filter Paper (GFFP) placed in a glass Petri-dish (GPD) of 9 cm internal diameter. Each GPD was moistened with 30 ml of apple of Sodom aqueous extract, sealed with parafilm, covered with black polyethylene bag, and incubated at 28°C in the dark. The treatments were arranged in completely randomized design with four replicates (r). The seeds were examined for germination at three days after initial germination. The percentage of the inhibition of seed germination was calculated using the following equation:

$$Inhibition (\%) = \frac{Total \ number \ of \ seeds - number \ of \ germinated \ seeds}{Total \ number \ of \ seeds} \times 100$$
(2)

# 5. Probit analysis procedure

Probit analysis was used to analyse data from bioassay experiment, *i.e.* the portions of seeds of African rattlebox inhibited by several concentrations of apple of Sodom. Results from probit analysis were reported typically as a concentration required inhibiting a certain portion of the test seeds (LC<sub>10</sub>, LC<sub>50</sub> and LC<sub>90</sub>); the slope and intercept of the regression line of the probit transformed data were also reported. Complementary probit transformed data were converted back to the concentration. In this study, probit analysis was achieved by conducting regression analysis, as follows:

Step 1: Correction of inhibition (%) using Abbott's formula

The inhibition (%) was corrected using Abbott's formula. It is given by:

Corrected inhibition (%) = 
$$\frac{X-Y}{X} \times 100$$
 (3)

Where:

X is the % survivorship of the control group (germination % in the control treatment).

Y is the % survivorship in the experimental group (germination % in the concentration treatment).

Step 2: Plotting of inhibition (%) against concentration (g/l)

Corrected inhibition (%) was plotted against concentration (%) to explore relationship between them.

Step 3: Transformation of data

The concentration (g/l) was transformed to log<sub>10</sub> concentration, (independent variable, X) and the corrected inhibition (%) was transformed to probits (dependent variable, Y) by using Finney's table [15], (Table-1).

%	0	1	2	3	4	5	6	7	8	9
0	-	2.67	2.95	3.12	3.25	3.36	3.45	3.52	3.59	3.66
10	3.72	3.77	3.82	3.87	3.92	3.96	4.01	4.05	4.08	4.12
20	4.16	4.19	4.23	4.26	4.29	4.33	4.36	4.39	4.42	4.45
30	4.48	4.50	4.53	4.56	4.59	4.61	4.64	4.67	4.69	4.72
40	4.75	4.77	4.80	4.82	4.85	4.87	4.90	4.92	4.95	4.97
50	5.00	5.03	5.05	5.08	5.10	5.13	5.15	5.18	5.20	5.23
60	5.25	5.28	5.31	5.33	5.36	5.39	5.41	5.44	5.47	5.50
70	5.52	5.55	5.58	5.61	5.64	5.67	5.71	5.74	5.77	5.81
80	5.84	5.88	5.92	5.95	5.99	6.04	6.08	6.13	6.18	6.23
90	6.28	6.34	6.41	6.48	6.55	6.64	6.75	6.88	7.05	7.33
-	0.0	0.1	0.2	0.3	0.4	0.5	0.6	0.7	0.8	0.9
99	7 33	7 37	7 41	7 46	7 51	7 58	7 65	7 75	7 88	8.09

**Table 1**-Table of transformation of percentages to probits, Finney's table

Step 4: Calculation of simple linear regression equation

The simple linear regression equation is: βX

$$Y = \alpha + \alpha$$

where:

Y: Probit value

X: Log<sub>10</sub> concentration

 $\alpha$ : intercept

 $\beta$ : regression coefficient, the slope

The simple linear regression equation was calculated as follows:

The regression coefficient, i.e. the slope value, was calculated by the following equation:

$$\beta = \frac{\sum_{i=1}^{n} X_i Y_i - \frac{\sum_{i=1}^{n} X_i \sum_{i=1}^{n} Y_i}{n}}{\sum_{i=1}^{n} X_i^2 - \frac{\left(\sum_{i=1}^{n} X_i\right)^2}{n}}$$
(5)

The intercept value was calculated by the following equation:

$$\alpha = \overline{Y} - \beta \overline{X} \tag{6}$$

Step 5: Testing the significance of the regression coefficient

To test the significance of the regression coefficient, i.e. the slope  $(\beta)$ , analysis of variance was conducted by testing the following hypotheses:

Null hypothesis:  $H_0: \beta = 0$ 

Alternative hypothesis:  $H_a: \beta \neq 0$ 

The analysis of variance depends on studying and partitioning the sum squires (SS) of the total to its basic components, regression sum squares, and error sum squares which were computed as follow:

(4)

$$SS.Total = \sum Y_i^2 - \frac{(\sum y_i)^2}{n}$$
(7)

$$SS.Regression = \frac{\left(\sum_{i=1}^{n} x_i Y_i - \frac{\sum_{i=1}^{n} x_i \sum_{i=1}^{n} Y_i}{n}\right)}{\sum_{i=1}^{n} x_i^2 - \frac{\left(\sum_{i=1}^{n} x_i\right)^2}{n}}$$
(8)

$$SS. Erorr = SS. Total - SS. Regression$$
(9)

Degree of freedom for total =
$$n-1$$
 (10)

$$Degree of freedom \ regression = I$$
(11)  

$$Degree of freedom \ error= DF \ for \ total- DF \ for \ regression$$
(12)

$$SS for regression$$
(12)

$$Mean squares for regression = \frac{Degree of freedom for rgression}{Degree of freedom for rgression}$$
(13)  

$$Mean squares for error = \frac{SS for Error}{Degree of freedom for rgression}$$
(14)

$$lean squares for error = \frac{BB for error}{DF for error}$$
(14)

$$F - cal \ for \ regression = \frac{MS \ for \ regression}{MS \ for \ error}$$
(15)

(16)

F-tab was obtained from F-table (F – Distribution).

The calculation processes of the analysis of variance were summarized in the ANOVA table. Step 6: Coefficient of simple determination  $(r^2)$ 

The coefficient of simple determination was calculated as follows:

$$r^2 = \frac{SS.Regression}{SS.Total}$$

Step 7: Calculation of the lethal concentrations

The LC<sub>25</sub>, LC<sub>50</sub> and LC<sub>75</sub> were calculated by substituting Y by 2.5, 5 or 7.5 in the regression equation (4), respectively. Then, the estimated log concentration was converted back (the antilog) to concentration.

#### 6. Statistical analysis

Step 1: Correction of inhibition (%) using Abbott's formula

The anticipated natural inhibition (%) was corrected using Abbott's formula (3) as shown in Table-2. E.g. Inhibition (%) and corrected inhibition (%) for the concentration 2.3 g/l were calculated as follows:

Inhibition (%) = 
$$\frac{100-95.75}{100} \times 100 = 4.3$$
 (2)

Corrected inhibition (%) = 
$$\frac{98.5 - 95.75}{98.5} \times 100 = 2.8$$
 (3)

**Table 2-**Concentration (g/l) of the leaves aqueous extract of apple of Sodom, inhibition (%), and correction of inhibition (%) of the seed germination of the African rattlebox using Abbott's formula

Concentration	Total number of	Number of	Inhibition (%)	Corrected
(g/l)	seeds	germinated seeds		inhibition (%)
0.0	100	98.50	1.5	0
2.3	100	95.75	4.3	2.8
4.6	100	90.50	9.5	8.1
7.0	100	75.50	24.5	23.4
9.3	100	50.75	49.3	48.5
11.6	100	34.75	65.3	64.8
13.9	100	20.75	79.3	79.0
16.2	100	10.50	89.5	89.3
18.5	100	6.00	94	93.9
20.8	100	4.00	96	95.9
23.2	100	1.75	98.3	98.3

Step 2: Plotting of inhibition (%) against concentration (g/l)

Corrected inhibition (%) was plotted against concentration (g/l) as shown in Figure-1.



Figure 1-Plotting corrected inhibition against concentration (%).

# Step 3: Transformation of data

Concentration (g/l) was transformed to  $log_{10}$ -concentration (X) and corrected inhibition (%) was transformed to probit (Y), as shown in Table-3.

E.g. at the concentration 9.3 g/l, the corrected inhibition (48.5  $\% \approx 49 \%$ ) was transformed to probits (4.97) by using Finney's table, as shown in Table-1.

Table 1-Table	of transformation	of percentages to	probits, Finney's table
		1 0	

%	0	1	2	3	4	5	6	7	8	9
0	-	2.67	2.95	3.12	3.25	3.36	3.45	3.52	3.59	3.66
10	3.72	3.77	3.82	3.87	3.92	3.96	4.01	4.05	4.08	4.12
20	4.16	4.19	4.23	4.26	4.29	4.33	4.36	4.39	4.42	4.45
30	4.48	4.50	4.53	4.56	4.59	4.61	4.64	4.67	4.69	4.72
40	4.75	4.77	4.80	4.82	4.85	4.87	4.90	4.92	4.95	4.97
50	5.00	5.03	5.05	5.08	5.10	5.13	5.15	5.18	5.20	5.23
60	5.25	5.28	5.31	5.33	5.36	5.39	5.41	5.44	5.47	5.50
70	5.52	5.55	5.58	5.61	5.64	5.67	5.71	5.74	5.77	5.81
80	5.84	5.88	5.92	5.95	5.99	6.04	6.08	6.13	6.18	6.23
90	6.28	6.34	6.41	6.48	6.55	6.64	6.75	6.88	7.05	7.33
-	0.0	0.1	0.2	0.3	0.4	0.5	0.6	0.7	0.8	0.9
99	7.33	7.37	7.41	7.46	7.51	7.58	7.65	7.75	7.88	8.09

Table 3-Transformation of concentration	to $\log_{10}$	concentration	(g/l) and	l corrected	inhibition	(%) to
probits using Finney's table.						

$Log_{10}$ concentration (X)	Probit (Y)
0.362	3.12
0.663	3.59
0.845	4.26
0.968	4.97
1.064	5.39
1.143	5.81
1.210	6.23
1.267	6.55
1.318	6.75
1.365	7.05

Probit was plotted against log<sub>10</sub> concentration (Figure-2).



Figure 2-Plotting probits against log<sub>10</sub> concentration.

Step 4: Calculation of the simple linear regression equation

α

The regression coefficient, i.e. the slope, value was calculated as follows:

$$\beta = \frac{\frac{58.653 - \frac{10.205 + 53.72}{10}}{11.330 - \frac{(10.205)^2}{10}} = 4.184$$
(5)

The intercept value was calculated as follows:

$$= 5.372 - 4.184 * 1.021 = 1.009 \tag{6}$$

The simple linear regression equation was:

$$Y = 1.099 + 4.184X \tag{4}$$

or

 $Probit = 1.099 + 4.184 \log_{10} concentration$ 

Step 5: Testing the significance of the regression coefficient

The significance of the regression coefficient was tested as follows:

$$SS.Total = 305.260 - \frac{(53.72)^2}{10} = 16.676$$
(7)

$$SS. Regression = \frac{\left(\frac{58.653 - \frac{10.205 \times 12}{10}}{11.330 - \frac{(10.205)^2}{10}}\right)}{11.330 - \frac{(10.205)^2}{10}} = 16.014$$
(8)

$$SS.Erorr = SS.Total - SS.Regression$$

$$SS. Erorr = 16.676 - 16.014 = 0.662 \tag{9}$$

$$Degree of freedom for total = 10-1=9$$
(10)

$$Degree of freedom \ regression = 1 \tag{11}$$

$$Degree of freedom \ error=9-1=8$$
(12)

Mean sqares for regression = 
$$\frac{10011}{1}$$
 = 16.014 (13)

Mean sqares for error 
$$=\frac{0.662}{\frac{8}{16}}=0.083$$
 (14)

$$F - cal \ for \ regression = \frac{16.014}{0.083} = 192.940 \tag{15}$$

F-tab values (5.5914 and 12.2466) were obtained from the F-table at alpha = 0.05 and 0.01, respectively.

The calculation processes of the analysis of variance are summarized in the ANOVA table (Table- 4).

		aa		<b>F</b> 1		F-Tab			
SOV	DF	SS	MS	F-cal	0.0	5	0.01		
Regression	1	14.014	16.014	192.940 **	5.31	77	11.2586		
Error	8	0.662	0.083						
Total	9	16.676							
Step 6: Coefficient o	f simple d	eterminati	$lon(r^2)$						
The coefficient of	simple det	ermination	n was calculated as	follows:					
	r <sup>2</sup>	$=\frac{14.014}{16.014}$	= 0.9603				(16)		
Step 7: Calculation c	of the letha	16.676 al concentr	ation						
$LC_{25} =$									
2.5 = 1.099 + 4.184	$4Log_{10}con$	ncentrati	ion						
•	2.5 -	- 1.099	0.00 <b>.</b>						
$Log_{10}$ concentratio	pn =	=	0.335						
Concentration = Ant	ilog of 0.3	35 = 2.16							
$LC_{50} =$									
5 = 1.099 + 4.184	Log <sub>10</sub> con	icentratio	on						
I a concentratio	$\frac{5-1}{2}$	1.099 – 0	032						
Log <sub>10</sub> concentration	$\frac{7}{4.1}$	184 <u>- 0</u>							
Concentration = Ant	ilog of 0.9	932 = 8.55							
$LC_{75} =$									
7.5 = 1.099 + 4.184	4 Log <sub>10</sub> co 7 r	ncentrat	ion						
$Log_{10}$ concentratio	$n = \frac{7.5 - 1}{4}$	$\frac{104}{104} =$	1.530						
Concentration = Ant	4. ilog of 1.8	$104 \\ 888 = 33.83$	8						
The calculation p	rocesses o	f the statis	stical analysis are su	mmarized in	Table-5.				
Table 5-Summary of	f the calcu	lation pro	cesses of the statistic	cal analysis.					
Concentration (g/l)	Corr	rected	Log <sub>10</sub>	Probit	$\mathbf{V}*\mathbf{V}$	$\mathbf{v}^2$	$\mathbf{v}^2$		
	inhibit	tion (%)	concentration (X)	(Y)	$\Lambda^{+}1$	Λ	1		
2.3	2	2.8	0.362	3.12	1.129	0.131	9.734		
4.6	8	3.1	0.663	3.59	2.380	0.440	12.888		
7.0	2	3.4	0.845	4.26	3.600	0.714	18.148		
9.3	4	8.5	0.968	4.97	4.811	0.937	24.701		
11.6	6	4.8	1.064	5.39	5.735	1.132	29.052		
13.9	7	9.0	1.143	5.81	6.641	1.306	33.756		

Table	4-ANOVA	table
Lanc		table

# **Results and Discussion**

X

16.2

18.5

20.8

23.2

89.3

93.9

95.9

98.3

The results showed that the leaves aqueous extract of apple of Sodom inhibited the seed germination of the African rattlebox with a direct positive relationship between concentration (g/l) and inhibition (%) (Table-2). Also, the results indicated a natural inhibition (%) in the seed germination of African rattlebox, as anticipated. Hence, before proceeding to conduct probit analysis, the inhibition (%) was corrected using Abbott's formula (Table-2). Plotting of corrected inhibition (%) against concentration (%) formed a sigmoid curve (Figure-1). Transformations of concentration to  $log_{10}$  concentration straightened the cumulative distribution line and the curve was transformed to more accurately describe the data (Figure-2).

1.210

1.267

1.318

1.365

10.205

1.021

6.23

6.55

6.75

7.05

53.72

5.372

7.538

8.299

8.897

9.623

58.65

1.464

1.605

1.737

1.863

11.33

38.813

42.903

45.563

49.703

305.26

The regression coefficient and the intercept values were 4.184 and 1.009, respectively. Hence, the simple linear regression equation was = 1.099 + 4.184X, *i.e. Probit* =  $1.099 * 4.184 \log_{10} concentration$ . Since the f-calculated value for regression was greater than the f-tabulated value at the levels of significance of 0.05 and 0.01, the regression coefficient was significantly different from zero. Therefore, the regression coefficient was included in the equation of simple linear regression. The value of coefficient of simple determination was 0.9603, i.e. 96.03% of the changes in the dependent variable y (inhibition) were caused by the independent variable x (concentration). The LC<sub>25</sub>, LC<sub>50</sub> and LC<sub>75</sub> were 2.16, 8.55 and 33.88 g/l, respectively.

Gulzar and Siddiqui [10] carried out an experiment to explore the effects of the aqueous extract of apple of Sodom on the seed germination of *Brassica oleracea*. Seeds of *B. oleracea* were steeped in solutions containing concentrations (20%, 40%, 60% and 80%) of leaves aqueous extract of apple of Sodom. The results revealed that the higher concentrations (60% and 80%) of the aqueous extract significantly reduced seed germination in comparison to the untreated control. The inhibitory effect was increased with the increase in the concentration of the aqueous extract. Gulzar and Siddiqui [10] concluded that the delayed germination and low germination rate of the *B. oleracea* after treatment by the aqueous extracts occurred because the extracts might damage the membrane system of the seeds.

The various concentrations of the leaves aqueous extracts of apple of Sodom had varying degrees of inhibition on the seed germination of mustard plant (*Brassica nigra*). The inhibition of seed germination was concentration-dependent. Suppression in the seed germination as a result of allelochemical stress might be attributed to inhibition of some physiological processes, such as water uptake, gibberellic acid activity, cell division, and elongation during germination process [16, 17].

The leaves aqueous extract of apple of Sodom at 5- 60 % has allelopathic effects on the seed germination of several crops such as barley (*Hordeum vulgare* L.), wheat (*Triticum aestivum* L.), cucumber (*Cucumis sativus* L.), fenugreek (*Trigonella foenum-graecum* L.), alssana (*Senna occidentalis* L.), tomato (*Lycopersicon esculentum* Mill.) and eggplant (*Solanum melongena* L.). The germination percentage was reduced with further increase in concentration of the aqueous extract [18, 19]. The delay in seed germination and the reduction in germination index might be due to the presence of water-soluble inhibitors in the apple of Sodom extract [9].

# Conclusions

The results revealed the effects of the leaves aqueous extract of apple of Sodom allelopathic on seed germination of African rattlebox, with a direct positive relationship between concentration (g/l) and inhibition (%). Also, the data indicated that plotting of corrected inhibition (%) against concentration (%) formed a sigmoid curve. Probit analysis transformed the sigmoid concentration-response curve to a straight line. Hence, the LC<sub>25</sub> (2.16 g/l), LC<sub>50</sub> (8.55 g/l) and LC<sub>75</sub> (33.88 g/l) were accurately estimated. Therefore, probit analysis is appropriate in studying the allelopathy phenomenon.

# References

- 1. Bliss C. I. 1934. The method of probits. *Science*, 79(2037): 38–39.
- 2. Simon, J. Yu. 2014. *The Toxicology and Biochemistry of Insecticides*. 2<sup>nd</sup> edition, CRC Press, Poca Raton, USA.
- **3.** Delabys, N., Mermillod G., De Joffrey J. P. and Bohren C. **2004**. Demonstration, in cultivated fields, of the reality of the phenomenon of Allelopathy. *XII. International conference on weed biology*, 97-104.
- 4. Inderjit, S. T. R, Callaway, R. M. and Vivanco, J. M. 2006. Can plant biochemistry contribute to understanding of invasion ecology?. *Trends plant science*. **11**: 1360–1385.
- 5. Singh, P. A. and Chaudharv, B. R. 2011. Allelopathic potential of algae weed Pithophora oedogonia (Mont.) ittrock on the germination and seedling growth of Oryza sativa L. *Botany Research International*, 4(2): 36-40.
- 6. [6] Gholami, B. A., Faravani, M. and Kashki, M. T. 2011. Allelopathic effects of aqueous extract from *Artemisia kopetdaghensis* and *Satureja hortensison* growth and seed germination of weeds. *Journal of Applied Environmental and Biological Sciences*, 1(9): 283-290.
- 7. Hussain, I. M. and Reigosa, M. J. 2011. Allelochemical stress inhibits growth, leaf water relations, PSII photochemistry, non-photochemical fluorescence quenching, and heat energy dissipation in three C3 perennial species. *Journal of Experimental Botany*, 62(13): 4533-4545.

- 8. Hassan, L. M., Galal, T. M., Farahat, E. A. and El-Midan, M. M. 2015. The biology of *Calotropis* procera (Aiton) W.T. *Trees*, 29: 311–320.
- Yasin, M., Safdar., M. E., Iqbal, Z., Ali, A., Jabran, K. and Tanveer, A. 2012. Phytotoxic effects of *Calotropis procera* extract on germination and seedling vogor of wheat. *Pak J Weed Sci Res*, 18: 379–392.
- **10.** Gulzar, A. and Siddiqui, M. B. **2017**. Allelopathic effect of *Calotropis procera* (Ait.) R. Br. on growth and antioxidant activity of *Brassica oleracea* var. botrytis. *Journal of the Saudi Society of Agricultural Sciences*, **16**: 375–382.
- Boatwright, J.S., Roux, M.M., Wink, M., andvan Wyk, B. 2008. Phylogenetic Relationships of Tribe Crotalarieae (Fabaceae) Inferred from DNA Sequences and Morphology. *Systematic Botany*, 33(4): 752-761.
- 12. Shah, A. B. S., Ud-Deen1, M. M., Naz. A, Sarker, J. K. and Kabir, G. 2008. Post-Irradiation Ageing Effect on Morphological Characters of *Crotalaria Saltiana*. J. bio-Sci. 16: 89-93.
- 13. Barri, M.E. S. and Adam, S. E. I. 1981. The Toxicity of *Crotalaria saltiana* to Calves. *J. of Comp. Path.* 91: 621-627.
- 14. Dafalla, G. A. and Cousin, M. T. 1988. Phyllody Disease of *Crotalaria saltiana* in the Sudan: Transmission to *Catharanthus roseus* and Detection of MLOs by Fluorescence and Electron Microscopy. *Journal of Phytopathology*, 123(3): 273-283.
- **15.** Randhawa, M. A. **2009**. Calculation of LD<sub>50</sub> values from the method of Miller and Tainter, 1944. *J Ayub Med Coll Abbottabad*, **21**: 184 –5.
- **16.** Olofsdotter, M. **1998**. Allelopathy in rice. In: Olofsdotter, M. (Ed.). Proceeding of the Workshop on Allelopathy in Rice, 25–27 November 1996. Manila, Philippines, International Rice Research Institute.
- 17. Tawaha, A.M. and Turk, M.A. 2003. Allelopathic effects of black mustard (*Brassica nigra*) on germination and growth of wild barley (*Hordeum spontaneum*). J. Agron. Crop Sci, 189: 298–303.
- **18.** Al-Zahrani, H. S. and Al-Robai, S. A. **2007**. Allelopathic effect of *Calotropis procera* leaves extract on seed germination of some plants. *J king Abdullah U*, **19**: 115–126.
- **19.** Ghasemi, S., Ghasemi, M., Moradi, N. and Shamili, A. M. **2012**. Effect of *Calotropis procera* leaf extract on seed germination of some plants. *JOHP*, **2**(1): 27–32.