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A Review of Entomopathogenic Fungi of Iraq

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Abstract

Insect pest management has been dominated by the use of synthetic pesticides since early 1950s. However, lately this control method is not widely accepted due to an increase in environmental awareness, food safety concerns and the increasing number of insecticide-resistant species. In Iraq, the chemical insect pest control is still a dominant control method regardless of the increased pressure to replace it gradually with environment friendly alternatives such as predators, parasitoids, nematodes and entomopathogenic fungi. In Iraq, there is an increasing volume of research that has used different genus and species of entomopathogenic fungi for controlling several agricultural pests. However, these efforts are not yet reflected in the industry. In this short review article, a detailed summary of the research that has been done with entomopathogenic fungi in Iraq since 2000 is given and a brief discussion of the potential of using entomopathogenic fungi on a large scale in Iraq, its challenges, and recommendations is presented.

Keywords: Entomopathogenic fungi, Iraq, Insect pest management, Biological control

مراجعة للفطريات الممرضة للحشرات في العراق

سنا نجم عبد الحديدي

قسم علوم الحياة، كلية العلوم، جامعة ديالى، ديالى، العراق

الخلاصة

لقد سيطر استخدام مبيدات الآفات الاصطناعية على مكافحة الآفات الحشرية منذ أوائل الخمسينيات من القرن الماضي، ومع ذلك في الآونة الأخيرة، أصبحت طريقة المكافحة هذه مرفوضة على نطاق واسع بسبب زيادة الوعي البيئي، وتنامي المخاوف المتعلقة بسلامة الغذاء وكذلك تزايد عدد أنواع الحشرات المقاومة للمبيدات الحشرية، أما في العراق فلا تزال المكافحة الكيميائية للآفات الحشرية هي طريقة المكافحة السائدة بغض النظر عن الضغط المتزايد لاستبدالها تدريجياً ببدائل صديقة للبيئة مثل المفترسات والطفيليات والديدان الخيطية والفطريات الممرضة للحشرات، مع العلم ان في العراق هناك عدد كبير من الأبحاث التي تستخدم أنواعاً مختلفة من الفطريات الممرضة للحشرات لمكافحة العديد من الآفات الزراعية، ومع ذلك، لم تنعكس هذه الجهود بعد في مجال مكافحة الآفات الحشرية على أرض الواقع، تضمنت مقالة المراجعة القصيرة هذه ملخص مفصل للبحوث التي تم إجرائها على الفطريات الممرضة للحشرات منذ سنة 2000 في العراق ،

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وكذلك تم مناقشة امكانية استخدام الفطريات الممرضة للحشرات في العراق على نطاق واسع وتسهيل الضوء على اهم التحديات التي تواجه ذلك وعرض بعض التوصيات لمواجهة تلك التحديات.

Introduction

Entomopathogenic fungi (EPF) are fungal species that are naturally pathogenic to insects. These fungal pathogens play a crucial role in the insect population regulation [1]. The start of this field was probably in 1835 when an Italian entomologist Agostino Bassi, announced that a fungus, that would eventually be named *Beauveria bassiana* in his honor, was transmitted among the silkworm by contact and by infected food. This silkworm disease could have given the idea of using insect infecting fungi for the control of insect pest management [2]. However, the fundamental field trial for using EPF to control insects was done by a Russian microbiologist, Elie Metchnikoff in 1888. Metchnikoff's work led to a revolution in the Biological control programs in Europe and the United States by using fungi to control insect pests [3].

Massive rise in the human population has led to more food demand [4], which was the major reason for using pesticides to control pests [5]. Since early 1950s, synthetic insecticides have been widely used to control crop pests as they showed a significant increase in crop productivity [6]. However, the abundance and unregulated use of pesticides led to the emergence of many health and environmental issues [6]. Another imperfection of using pesticides is that a large number of insecticides affect non-target arthropod species. Moreover, insects begin to develop resistance to those synthetic pesticides that led to increased pesticide dosage in every new application [7]. All these issues have forced the industry and researchers to focus on the development of alternative control methods, including many biological control agents and EPF [6].

The production of EPF for commercial purposes is not very expensive. It does not require strict storage conditions as the storage can be efficient in a wide range of humidity and temperature. Therefore, EPF is being developed and produced on mass production globally to control a wide range of agricultural pests [8]. However, in Iraq, using biological control agents to control agricultural pests is not a prior option for the farmers. Thus, hundreds of Iraqi researches have been conducted in this area. Most of these studies have been published in Arabic and are, therefore, not readable internationally, and are isolated from being invested globally. So far no local commercial products are available in the Iraqi market. Hence, it is necessary to highlight, illustrate and discuss the progress that has been done in this field in Iraq to establish a database for more focused research in local EPF species for commercial purposes.

A summary of the EPF research in Iraq since 2000 is given in this short review article. A bibliography of the published articles with their URLs has been provided at the end to increase the visibility for these articles. A list of EPF species that have been experimentally tested, with the insect pest species that were tested against, is also provided. A brief classification of these species, their sources and experimental duration are also stated. Finally, the challenges of using and developing the EPF industry in Iraq are discussed, with some recommendations to encounter these challenges.

Summary of EPF Research in Iraq

The earliest known study on EPF in Iraq was in the early 1980s. However, the real interest in this subject in Iraq started at the beginning of the 21st century. It is important to state that the collected published articles for this review represent the accessible online work since 2000. There are 128 experiments within 117 published articles that tested EPF on different

arachnids' pests. Ninety-four of these experiments were done in labs, 27 in the field and 7 in greenhouses. The most experimentally tested species was *Beauveria bassiana* (Bals. -Criv.) Vuill., (99 experiments within 88 studies) followed by *Metarhizumanisopliae* (Metchnikoff) (27 experiments within 26 studies) (Table 1).

Total 30 species were experimentally tested EPF species (Table 1). Taxonomically, they belong to 16 genera (*Beauveria*, *Metarhizium*, *Lecanicillium*, *Chaetomium*, *Isaria*, *Penicillium*, *Cladosporium*, *Alternaria*, *Aspergillus*, *Mucor*, *Paecilomyces*, *Trichoderma*, *Scytalidium*, *Emericella*, *Mycelophthora*, *Fusarium*), 7 orders (Hypocreales, Sordariales, Eurotiales, Capnodiales, Pleosporales, Mucorales, Helotiales), and 5 classes (Sordariomycetes, Eurotiomycetes, Dothideomycetes, Leotiomycetes, Zygomycetes). 15 of these genera belong to Division Ascomycota and just one Genus (*Mucor*) belongs to Division Mucoromycota.

In these 117 published articles, there are 26 genera and 42 species of EPF isolated from infected insects, insects cadavers, soil and sewers of the Iraqi environment (Table 2). EPF was tested against 48 pest species (Table 1). 5 of these species belonged to class Arachnida (Orders: Ixodida and Trombidiformes), while 42 species belonged to class Insecta (Hemiptera, Lepidoptera, Coleoptera, Diptera, Blattodea, Hymenoptera, Isoptera, Orthoptera).

Table1: List of Entomopathogenic fungi species experimentally tested vs. insect pest species in Iraq and the experiment types.

EPF	Insect pest and their stages	Experiment type	Reference
<i>Beauveria bassiana</i> (Bals.-Criv.) Vuill.	Green peach aphid <i>Myzus persicae</i> (Sulzer) (different stages)	greenhouse	[9]
	Khapra Beetle <i>Trogoderma granarium</i> (Everts.) (eggs)	lab	[10]
	The Dust Spider Mite <i>Oligonychus afrasiaticus</i> (different stages)	lab and field	[11]
	Subterranean termite <i>Microcerotermes diversus</i>	field	[12]
	Cotton leaf worm <i>Spodoptera littoralis</i>	field	[13]
	Black bean aphid <i>Aphis fabae</i>	lab	[14]
	House fly <i>Musca domestica</i> (L.)	lab	[15]
	Jasmine whitefly <i>Aleuroclava jasmine</i> (egg and nymph)	field	[16]
	Angomis grain moth <i>Sitotrogacerealella</i> (Oliver) (larvae)	lab	[17]
	Cotton leaf worm <i>Spodoptera littoralis</i> (larvae)	lab	[18]
	German cockroach <i>Blattella germanica</i> (L.)	lab	[19]
	Termites <i>Microcerotermes diversus</i> (Silvestri)	lab and field	[20]
	Corn stem borer <i>Sesamiacretica</i> (Led.) (larvae)	lab and field	[21]
	Khapra beetle <i>Trogoderma granarium</i> (Everts) (larvae)	lab	[22]
	Great Moth Wax <i>Galleria mellonella</i> (L.) (larvae)	lab	[23]
	House fly <i>Musca domestica</i> (L.) (all stages)	lab	[24]
	Cowpea weevil <i>Callosobruchus maculatus</i> (Fab.) (all stages)	lab	[25]
	Potato tuber moth <i>Phthorimaea operculella</i>	lab and field	[26]
	Cotton leave worm <i>Agrotis ipsillia</i>	lab and field	[27]
	Whitefly <i>Bemisia tabaci</i> (nymph, adults)	field and greenhouse	[28]
	Brown banded cockroach <i>Supella longipalpa</i> (egg, adult)	lab	[29]
	Potato tuber moth <i>Phthorimaea operculella</i> (SELLER) (egg, larvae, pupa)	lab	[30]

Two spotted spider mites <i>Tetranychusurticae</i>	field	[31]
Broad mite <i>Polyphagotarsonemus latus</i> (Banks)	field	[32]
Wax moth <i>Galleria mellonella</i> (egg and larvae)	lab	[33]
Black bean aphid <i>Aphis fabae</i> (Scopolli)	lab and field	[34]
Termite <i>Microcerotermesdiversus</i> (Silvestri) (workers and soldiers)	lab	[35]
Wax moth <i>Galleria mellonella</i> (adult)	lab	[36]
Cotton whitefly <i>Bemisiatabaci</i> (nymphal stages)	greenhouse	[37]
Citrus mealybug <i>Planococcuscitri</i> (different stages)	lab	[38]
Onion maggot <i>Delia alliaria</i> (Fonseca)(different stages)	lab	[39]
Onion maggot <i>Delia alliaria</i> (Fonseca)	field	[40]
Alsdi <i>Triboliumcastaneum</i> (Herbst) (adult)	lab	[41]
Mealy plum aphid <i>Hyalopteruspruni</i> (Geoffroy) and <i>Aphis pomi</i> (DeGree)	lab	[42]
Moth <i>Ephestiacautella</i> (Walk.)(larvae)	lab	[43]
Sunn pest <i>Eurygasterintegriceps</i> (Put.) (eggs) and parasitoid <i>Trissolcus</i> spp. (eggs)	lab	[44]
Cowpea seed beetle <i>Callosobruchus maculatus</i> (Fab.) (larval stages and adult)	lab	[45]
Tomato leafminer <i>Tutaabsoluta</i> (eggs and larval stages)	lab	[46]
Lesser grain borer <i>Rhyzoperthadominica</i> (Fab.) (adult)	lab	[47]
Fig Leaf Worm <i>Ocnerogyiaamanda</i> (Stgr.)	field	[48]
Hard ticks <i>Boophilusmicroplus</i> (adult)	lab	[49]
Rust red beetle <i>Triboliumcastaneum</i>	lab	[50]
Sun pest <i>Eurygasterintegriceps</i> (Put)(adult)	lab	[51]
Sun pest <i>Eurygasterintegriceps</i> (Put)(adult)	field	[52]
<i>Triboliumcastaneum</i> (Herbst) (larval stages)	lab	[53]
Potato tuber moth <i>PhthorimaeaOperculella</i> (Zeller) (all stages)	lab	[54]
Onion maggot <i>Delia alliaria</i> (Fonseca)	field	[55]
White scale insect <i>Parlatoriablanchardi</i> (Targ.)	lab	[56]
Mediterranean fruit fly <i>Ceratitiscapitata</i> (larvae)	lab	[57]
Homaira insect <i>Batrachedraamydraula</i> (egg larval stages 1,5)	lab	[58]
Khapra beetle <i>Trogoderma granarium</i> (Everts)	lab	[59]
Rrusty flour beetle <i>Triboliumcastaneum</i> and <i>Rhyzoperthadominica</i> (Fab.) (different larval stages)	lab	[60]
Sugar beet worm <i>Spodoptera exigua</i> (Hübner)	lab and greenhouse	[61]
Fig moth <i>EphestiaCautella</i> (Walk.)(all stages)	lab	[62]
<i>Culex quinquefasciatus</i> (Say)(4 th larval stage)	lab	[63]
Mosquito <i>Culex pipienspipiens</i> L.(larve)	lab	[64]
Citrus mealybug <i>Planococcuscitri</i> (Risso)	lab	[65]
Fruit stalk borer <i>Oryctesspp.</i>	field	[66]
Lesser pumpkin fly <i>Dacusciliatus</i> Loew (adult)	lab	[67]
<i>Triboliumcastaneum</i> (Herbst) (adult)	lab	[41]
House Fly <i>Musca domestica</i> L.(Pupa and adult)	lab	[68]
Green peach aphid <i>Myzus persica</i> (sulzer)	lab	[69]
Two-spotted spider mite <i>Tetranychusurticae</i> (nymph and adults)	lab	[70]

<i>Metarhiziumanisopliae</i>	<i>Spodoptera littoralis</i> (Boisd.)	greenhouse	[71]
	White fly <i>Bemisia tabaci</i> (Gennadius)(nymphs and adults)	field	[72]
	Dubas Bug <i>Ommatissus lybicus</i> (Deberg)	field	[73]
	Mole cricket <i>Gryllotalpa</i> sp.	field	[74]
	German cockroach <i>Blattella germanica</i> (nymphs and adults)	lab	[75]
	Moth <i>Ephesiaca cautella</i> (larval stages)	lab	[76]
	Angoumois moth <i>Sitotrogacerealella</i> (different stages)	lab	[77]
	<i>Trogoderma granarium</i> (Herbst)(larvae)	lab	[78]
	Moth Figs <i>Ephesiaca cautella</i> (Walk.) (eggs and larvae)	lab	[79]
	Palm dubas bug <i>Ommatissus lybicus</i> (adults)	lab	[80]
	Citrus mealybug <i>Planococcus citri</i> (Risso)	greenhouse	[81]
	Black fly <i>Acaudalero desrachipora</i> (Singh) (all stages)	field	[82]
	Brown Banded Cockroach <i>Supella longipalpa</i> (F.) (different stages)	lab	[83]
	Red spider mite <i>Tetranychus urticae</i> (Koch.)	field	[84]
	Sunn pest insect <i>Eurygaster testudinaria</i> (Geofroy)	lab	[85]
	Tomato leaf minor <i>Tuta absoluta</i> (Meyrick)	lab	[86]
	<i>Culex quinquefasciatus</i> (larvae)	lab	[87]
	Corn Stem Borer <i>Sesamia cretica</i> (Led)	field	[88]
	Cucurbit fruit fly <i>Dacus ciliatus</i> (Leow) and Melon Fruit fly <i>Dacus frontalis</i> (Beker)	field	[89]
	Black bean aphid <i>Aphis fabae</i> (adult)	field	[90]
	Beetle <i>Epilachna chrysomelina</i> (larvae)	lab and field	[91]
	Cowpea weevil <i>Callosobruchus maculatus</i>	lab	[92]
	<i>Bagradahilaris</i> (Burmeister) (nymphs and adults)	lab and field	[93]
	Green peach aphid <i>Myzus persicae</i> (Sulzer) (adult)	lab and field	[94]
	Flour beetle <i>Tribolium castaneum</i> (adult)	lab	[95]
	<i>Culex pipiens pipiens</i>	lab	[96]
	<i>Callosobruchus maculatus</i>	lab	[97]
	Corn borer <i>Sesamia cretica</i> Led.	field and lab	[98]
	House fly <i>Musca domestica</i> L. (larvae)	lab	[99]
	House fly <i>Musca domestica</i> L. (all stages)	lab	[100]
	Termite <i>Microcerotermes diversus</i> (Silv.)	field	[101]
	Lesser grain borer <i>Rhyzopertha dominica</i> (Fab.) (adult)	lab	[47]
	Potato tuber moth <i>Phthorimaea operculella</i> (Zeller) (all stages)	lab	[54]
	<i>Culex quinquefasciatus</i> (Say) (all stages)	lab	[102]
	Mediterranean fruit fly <i>Ceratitis capitata</i> (prepupae larvae)	lab	[57]
	Cowpea beetle <i>Callosobruchus maculatus</i> Fab.	lab	[103]
	Sawtoothed grain beetle <i>Oryzaephilus surinamensis</i> L. (all stages)	lab	[104]
	<i>Culex quinquefasciatus</i> (Say) (4 th larval stage)	lab	[63]
	Fruit stalk borer <i>Oryctes</i> spp.	field	[66]
	<i>Chrysomya albiceps</i> (Wiedeman) (larvae and adults)	lab	[105]
	Lesser grain borer <i>Rhyzopertha dominica</i> (Fab.) (all stages)	lab	[106]
	Lesser pumpkin fly <i>Dacus ciliatus</i> Loew (adult)	lab	[67]
	Blow fly <i>Chrysomya albiceps</i> (Wiedemann) (larvae)	lab	[103]
	House Fly <i>Musca domestica</i> L. (Pupa and adult)	lab	[68]

Lecanicillium muscarium = Verticillium lecanii (Zimm.)	Mole cricket <i>Gryllotalpa</i> sp.	field	[74]
	Sunn pest insect <i>Eurygaster testudinaria</i> (Geofroy)	lab	[85]
	Dubas bug <i>Ommatissus lybicus</i> Bergevin (different stages)	lab	[108]
	Housefly <i>Musca domestica</i> (adult)	lab	[109]
	<i>Culex quinquefasciatus</i> (egg, larvae, pupa and adult)	lab	[110]
	Aphid <i>Myzus persicae</i> , <i>Aphis gossypii</i> , <i>Aphis fabae</i> and <i>Macrosiphum euphorbiae</i>	lab and greenhouse	[111]
	<i>Callosobruchus maculatus</i> (Fab.) (adults)	lab	[112]
	Hard ticks <i>Hyalomma anatolicum</i> (adults)	lab	[113]
	Grey scale insect <i>Parlatoria blanchardi</i>	lab and field	[114]
	German cockroach <i>Blattella germanica</i> (L.)	lab	[19]
	Corn borer <i>Sesamia cretica</i> Led.	field	[98]
	Whitefly <i>Bemisia tabaci</i> (nymph, adults)	field and greenhouse	[28]
	Potato tuber moth <i>Phthorimaea operculella</i>	lab and field	[26]
	Cotton leaf worm <i>Agrotis ipsillia</i>	lab and field	[27]
	Rust red beetle <i>Tribolium castaneum</i>	lab	[50]
	<i>Aphis craccivora</i> Koch (nymph and adult)	lab	[115]
	Moth <i>Ephesiaca cautella</i> (Walk.) (all stages)	lab	[116]
	Cucurbit fruit fly <i>Dacus ciliatus</i> (Leow) and Melon fruit fly <i>Dacus frontalis</i> (Beker)	field	[89]
	Aphid <i>Myzus persicae</i> , <i>Aphis gossypii</i> , <i>Aphis fabae</i> and <i>Macrosiphum euphorbiae</i>	lab and greenhouse	[111]
Termites <i>Microcerotermes diversus</i>	lab	[117]	
Two-spotted spider mite <i>Tetranychus urticae</i> (nymph and adults)	lab	[70]	
Aphid <i>Myzus persicae</i> , <i>Aphis gossypii</i> , <i>Aphis fabae</i> and <i>Macrosiphum euphorbiae</i>	lab and greenhouse	[111]	
<i>Myzus persicae</i> (Sulzer) (nymph and adults)	lab	[118]	
Mealy plum aphid <i>Hyalopterus pruni</i> (Geoffroy) and <i>Aphis pomi</i> (DeGree)	lab	[42]	
Mealy plum aphid <i>Hyalopterus pruni</i> (Geoffroy) and <i>Aphis pomi</i> (DeGree)	lab	[42]	
Black bean aphid <i>Aphis fabae</i> (Scopoli)	lab and field	[34]	
Termites <i>Microcerotermes diversus</i>		[117]	
German cockroach <i>Blattella germanica</i> (L.)		[19]	
<i>Periplaneta americana</i>		[119]	
Black bean aphid <i>Aphis fabae</i> (Scopoli)	lab and field	[34]	
Termites <i>Microcerotermes diversus</i>	lab	[117]	
Black bean aphid <i>Aphis fabae</i>	lab	[14]	
Black bean aphid <i>Aphis fabae</i> (Scopoli)	lab	[34]	
White scale insect <i>Parlatoria blanchardi</i> (Targ.)	lab	[56]	
Two-spotted spider mite <i>Tetranychus urticae</i> (nymph and adults)	lab	[70]	
<i>Periplaneta americana</i>	lab	[119]	
Termites <i>Microcerotermes diversus</i>	lab	[117]	

<i>flavus, A. fumigatus</i>			
<i>Aspergillus candidus, A. niger, A. flavus, A. ochraceus, A. terreus</i>	Black bean aphid <i>Aphis fabae</i>	lab	[14]
<i>Aspergillus sp.</i>	German cockroach <i>Blattella germanica</i> (L.)	lab	[19]
<i>A. niger</i>	Black bean aphid <i>Aphis fabae</i> (Scopoli)	lab and field	[34]
<i>Aspergillus terreus, A. oryzae, A. tritici,</i>	Two-spotted spider mite <i>Tetranychus urticae</i> (nymph and adults)	lab	[70]
<i>Aspergillus sp.</i>	Whitefly insect <i>Bemisia tabaci</i> Genn. (nymph and adult)	greenhouse	[120]
<i>Aspergillus niger, A. parasiticus</i>	<i>Periplaneta americana</i>	lab	[119]
<i>Aspergillus niger, A. fumigatus, A. fumigatus, Mucor sp., Mucor sp., Cladosporium sp.</i>	Termites <i>Microcerotermes diversus</i>	lab	[121]
<i>Paecilomyces variotii</i>			
<i>Trichoderma hamatum</i>			
<i>Scytalidium lignicola</i>			
<i>Trichoderma hamatum</i> (Rifia)	Termites <i>Microcerotermes diversus</i>		[117]
<i>Trichoderma harzianum</i> (Rifia)			[10, 14, 29, 48, 73, 84, 19]
<i>Trichoderma harzianum.</i>	Field cricket <i>Gryllodessa igillatus</i> (nymph and adult)	lab	[122]
<i>Emericella vericolla, Myceliophthora sp.</i>			[14]
<i>Fusarium solani</i>			[20, 56]
<i>Fusarium semitectum</i>	Wax worm larvae <i>Galleria mellonella</i>	Lab	[123]
<i>Fusarium sp.</i>			[14, 111]

Table 2: Entomopathogenic fungi recorder in Iraq and their sources

Entomopathogenic fungi	Source	Reference
<i>Mucor sp.</i>	Cadavers and infected termites	[121]
<i>Chaetomium globosum</i>	<i>Microcerotermes diversus</i> From palm groves in Basra	
<i>Alternaria alternata</i>	Governorate, Abi Al-Khasib District	
<i>Aspergillus niger</i>		
<i>A. flavus</i>		
<i>A. fumigatus</i>		
<i>Cladosporium sp.</i>		
<i>Paecilomyces variotii</i>		
<i>Penicillium sp.</i>		
<i>Scytalidium lignicola</i>		
<i>Trichoderma hamatum</i>		
<i>Beauveria bassiana</i> (different isolates)	Larvae of date palm stem-borer <i>Jebusaeahammerschmidti</i>	[9,12]
	<i>Spodoptera littoralis</i> on potato plant	[13]
	Cadavers of sunn pest insect <i>Eurygaster testudinaria</i> (Geofroy)	[27, 28, 44, 85]
	Soil samples from fields, forests and vine yard in Duhok Governorate	[42]
	Soil samples from agro-ecosystems from Baghdad, Wasit, Diyala, Salah AL-din, Babil, Karbala, Amarah,	[124]

	DhiQar, Al-Qadisiyyah, Al-Muthanna, Dohuk, Sulaymaniyah and Erbil	
	Soil samples from palm and citrus orchards Baghdad (Koerich and Numaniyah)	[63]
	Soil samples from fields in Baghdad and citrus orchards in Kut	[67]
	Two isolates from date palm, <i>Phoenix dactylifera</i> L. leaves, and one from soil Al-Mada'in district/ Baghdad.	[73]
	Black fly <i>Acaudalerodesrachimpora</i> Singh on Sidr trees	[82]
	Soil samples from Gara Mountain/ Duhok	[91]
<i>Beauveria varroa</i> (first record)	Soil samples from the hibernation site of sunn pest	[91, 125]
<i>Beauveria pseudobassiana</i> Rehner and Humber (first record at 2019)	<i>Eurygasterintegriceps</i> Puton at Gara Mountain, Duhok	
<i>Isaria javanica</i> (first record)	Soil samples from fields, forests and vine yard in Duhok Governorate	[42]
<i>Penicillium glabrum</i>	Black Bean Aphid <i>Aphis fabae</i> (Scopoli)	[14]
<i>Alternaria alternata</i> , <i>Aspergillus candidus</i> , <i>A. niger</i> , <i>A. flavus</i> , <i>A. ochraceus</i> , <i>A. terreus</i>		
<i>Emericella vermicolla</i> , <i>Fusarium</i> sp., <i>Myceliophthora</i> sp., <i>Trichoderma harzianum</i>		
<i>Verticillium lecanii</i>	Whitefly <i>Bemisia tabaci</i>	[27, 28]
	Corn borer <i>Sesamia cretica</i> Led. on sorghum crop	[98]
	Soil samples from agro-ecosystems from Baghdad, Wasit, Diyala, Salah AL-din, Babil, Karbala, Amarah, DhiQar, Al-Qadisiyyah, Al-Muthanna, Dohuk, Sulaymaniyah and Erbil	[124]
<i>Alternaria alternata</i> , <i>A. niger</i> , <i>Beauveria bassiana</i> , <i>Cladosporium oxysporum</i> , <i>Penicillium chrysogenum</i> , <i>P. compactum</i> , <i>Ulocladium atrum</i> , <i>P. chlamydospora</i>	Black aphid <i>Aphis fabae</i>	[34]
<i>Alternaria alternate</i> , <i>A. dianthicola</i> , <i>Cladosporium herbarum</i> , <i>C. oxysporum</i> , <i>Fusarium solani</i> , <i>Helminthosporium</i> sp., <i>Phoma glomerata</i> , <i>Stemphylium</i> sp., <i>Ulocladium</i> sp., <i>Beauveria bassiana</i>	White scale insect <i>Parlatoria blanchardi</i> Targ. from three cites from Basra Governorate (Garmat Ali, Shatt al-Arab and the city center)	[56]
<i>Paecilomyces farinosus</i> (Holm.) <i>Paecilomyces lilacinus</i>	Cadavers of sunn pest	[44]
	Soil samples from agro-ecosystems from Baghdad, Wasit, Diyala, Salah AL-din, Babil, Karbala, Amarah, DhiQar, Al-Qadisiyyah, Al-Muthanna, Dohuk, Sulaymaniyah and Erbil	[124]
<i>Metarhiziumanisopliae</i> (different strains)	Soil samples from palm and citrus orchards from Baghdad/ Numaniyah and Basra/ Abu Khasib	[63]
	Soil samples from fields /Baghdad and citrus orchards /Kut	[67]
	Cadavers of sunn pest insect <i>Eurygaster testudinaria</i> (Geofroy)	[85]
	Corn borer <i>Sesamia cretica</i> Led. on sorghum crop	[98]
	Cadavers of <i>Culex quinquefasciatus</i>	[110]
	Larvae from Sewers of Al-Qadisiyah city	
	Soil samples from agro-ecosystems from Baghdad, Wasit, Diyala, Salah AL-din, Babil, Karbala, Amarah, DhiQar, Al-Qadisiyyah, Al-Muthanna, Dohuk,	[124]

<i>Beauveria bassiana</i> , <i>Pithomycescharatarum</i> , <i>Aspergillus terreus</i> , <i>A. oryzae</i> , <i>A. tritici</i> , <i>Aspergillus sp.</i> <i>Alternaria alternata</i> , <i>Chaetomium globosum</i> <i>Beauveria bassiana</i> , <i>Asperigllusparasiticus</i> , <i>A. flaves</i> <i>Cladosporium sp.</i> <i>Phoma glomerata</i> <i>Fusarium semitectum</i> (first record) <i>Fusarium sp.</i> <i>Aspergillus flavus</i> , <i>A. niger</i> <i>Cunninghamellaechinulata</i> , <i>Emericellanidulans</i> , <i>Fusarium spp.</i> , <i>Mucor spp.</i> , <i>Penicillium sp.</i> <i>Rhizopus stolonifer</i> <i>Trichoderma sp.</i> <i>Lecanicilliumlecanii</i> <i>Metarhiziumanisopliae</i> (first record from Iraqi soil) <i>Aspergillus niger</i> , <i>A. parasiticus</i> , <i>Aspergillus niger</i> <i>A.fumigatus</i> <i>Trichoderma harzianum</i> <i>Penicillium sp.</i> , <i>Alternaria sp.</i> , <i>Mucor sp.</i> , <i>Fusariumsp.</i>	Sulaymaniyah and Erbil	
	Two-spotted spider mite on egg plant	[70]
	Moth <i>Ephestiacautella</i>	[76]
	Fruit stalk borer <i>Oryctes elegans</i> larvae from Basra governorate	[123]
	Soil samples from agro-ecosystems from Baghdad, Wasit, Diyala, Salah AL-din, Babil, Karbala, Amarah, DhiQar, Al-Qadisiyyah, Al-Muthanna, Dohuk, Sulaymaniyah and Erbil	[124]
	Soil samples from different agro ecosystems at Duhok governorate (Alfalfa fields, different vegetables, forest of the endemic Pine <i>Pinus brutia</i> (Ten.), and soil of figs, berry, pear and walnut)	[126]
	Cadavers and honeydew of <i>Bemisiatabaci</i> Genn	[120]
	Women's and obstetricians hospital and residential areas near of the hospital/ Kerbala	[119]

Sources of EPF Tested Isolates and Experimental Durations

EPF used in these studies were either isolated from infected or dead insects (Table 2) such as larvae of *Jebusaeahammerschmidt* [9, 12], *Ephestiacautella* moth [76] black flies *Acaudalerodesrachipora* [82] *Spodoptera littoralis* [13], *Aphis fabae* [34] and sunn insect *Eurygaster testudinaria* [26, 27, 28, 44, 51, 52, 85], or were isolated from agricultural soils [42, 63] and sewers [110]. Some of the used EPF were obtained from research centres or universities of other Arab countries such as Jordan [43] and Egypt [41, 99, 100]. A big portion of the used EPF was commercial formulation such as Mycotal from Koppert, Netherlands [38, 50], Naturalis from Troy- Bioscience –Arizona, USA [30, 35, 47], Biocont from Dow Agrosiemces, USA [48,7 4] (ORY-X) from Felda Agriculture, Malaysia [74, 101], Recharge from Russell IPM company, UK [89]. While most of the other used EPF were obtained from lab strains conserved either at universities or research centres in Iraq.

Experiments durations varied. Lab and greenhouse experiments ranged between 2-21 days while field experiments, in general, took a bit longer period and reached up to 14 weeks [12]. In general, all the experiments showed satisfying to excellent results in controlling pests, without any study indicating a failure in their outcomes.

The Status of Using EPF in Iraq, its Challenges, and Recommendations

There is an increased demand for safe, eco-friendly products globally, and using EPF and other biological control products to control agricultural pests that fulfils this demand [4].

These products reduce pesticide use, minimize the negative impact of their use on the environment and guarantee farmers safety while maintaining crop production [6, 127]. In many countries, an entire industry has developed to produce, disseminate and aid in the adoption of natural enemies use [7, 8, 9]. This came up with a new generation of growers who are more concerned about pesticide use, its potential non-target effects, and other environmental and health issues. Supported by the government legislation and rules that provide the foundation for regulation of the introducing and applying natural enemies of insects as biological control agents [127]. In Iraq, despite all the above-listed work that has been done in this area, no local commercial products have emerged from this effort. Many challenges are faced in the establishment of this industry in Iraq. There is not enough awareness among the public and farmers towards the side effects of using chemical pesticides to control pests. Thus, increasing farmers' awareness of the environment and health issues associated with using chemical pesticides, could be a good starting point. Financial investment in research to develop this industry and the commercial interests are absent. Therefore, a long-term government support is required to support more focused research in Universities, research centres and the Ministry of Agriculture. Changing the chemical culture that Iraqi farmers and governments approaching for decades needs a genuine change in government attitude in controlling agriculture pests. Led by emphasizing the importance of preserving these species for the environment in education, especially in the faculties of agriculture and media, is needed. Following that proper technology is provided to support this approach, proper legislation is needed to implement rules and regulations that organise the importing, breeding, transporting and applying of these organisms. Also, there is a need for International cooperation represented by transporting experience and providing essential support techniques.

Conclusion

There is a global movement toward replacing chemical pesticides with eco-friendly controlling pest methods and Iraq has to follow this movement. Although the volume of research that has been done by Iraqi researchers to investigate and test the applicability of using different organisms such as EPF to control agricultural pests, there is no satisfying progress in this field yet. There is a dire need for more focused research, necessary technology, successful marketing, farmers' awareness, international cooperation and government support to achieve this goal.

Conflict of interest

The author declared no conflict of interest to this review article.

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