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## Histopathological and Biochemical Study on the Kidneys of Male Mice Injected Intraperitoneally with Thioacetamide

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### Abstract

Thioacetamide (TAA) is a thiono-sulfur containing compound with a wide manufacturing application. Humans and animals exposure to TAA may occur in different ways and may cause nephrotoxicity. So, in this study serum creatinine concentration and urea, in addition to renal pathological changes, were examined in mice treated with TAA/P (100 mg/kg B.W). One hundred twenty male albino (BALB/c) mice were used. They were randomly divided into 2 main groups. In the control group 30 mice were fed water only and normal mice pellet. The other TAA-treated group of mice were divided into 3 subgroups as follow: 1<sup>st</sup> group (G1) was injected with TAA for 2 months (injected twice a month), while the 2<sup>nd</sup>(G2) and 3<sup>rd</sup>(G3) groups were injected with TAA for 4 and 6 months respectively (single injection monthly). Subsequently, serum urea and creatinine were measured in the control and the treated mice. The pathological changes in renal tissue were studied by examiningstained Hematoxylin and eosin (H and E). The results showed a significant increase ( $P<0.05$ ) in blood urea and creatinine levels of all studied groups with diverse pathological changes in kidney, including degeneration, necrosis, hemorrhage, nephritis, abscess and granulomatous lesions due to TAA toxicity.

**Keywords:** TAA, Urea, Creatinine, Necrosis, Renal failure.

### دراسة نسيجية مرضية و كيميائية حيوية على كلى ذكور الفئران المحقونة داخل الصفاق بالثيواسيتاميد

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### الخلاصة

الثيواسيتاميد هو مركب الكبريت العضوي والذي يدخل في مجال الصناعة وله العديد من التطبيقات. لذلك من الممكن ان يتعرض الإنسان والحيوان لهذه المادة بطرق مختلفة وقد ينتج عن ذلك تسمم في الكلى. تناولت هذه الدراسة قياس مستوى تركيز الكرياتينين واليوريا في دم الفئران التي تم معاملتها بالمركب بتركيز 100 ملغم لكل كغم من وزن الفأر بالإضافة إلى دراسة التغيرات التي حدثت في الانسجة الكلوية. قسمت الفئران والبالغ عددها 120 بشكل عشوائي إلى مجموعتين رئيسيتين: مجموعة السيطرة والبالغ عددها 30 فأراً تلقت الماء و الغذاء الاعتيادي فقط ، المجموعة الأخرى والتي تم معاملتها بالثيواسيتاميد مكونة من 90 فأراً و مقسمة إلى 3 مجموعات و على النحو التالي: المجموعة الأولى (G1) والتيتم حقنها ب TAA لمدة

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شهرين (مرتين في الشهر) ، بينما المجموعة الثانية (G2) والمجموعة الثالثة (G3) تم حقنها بـ TAA لمدة 4 و 6 أشهر على التوالي (حقنة واحدة في الشهر). بعد ذلك ، تم جمع عينات المصل في الشهر الثاني والرابع والسادس عن طريق ثقب القلب لغرض قياس مستوى اليوريا والكرياتينين قبل وبعد المعاملة. كذلك تمت دراسة التغيرات المرضية في النسيج الكلوية من خلال فحص عينات الأنسجة المصبغة بصبغة الهيماتوكسيلين والايوسين. أظهرت النتائج زيادة معنوية ( $P < 0.05$ ) في مستوى اليوريا والكرياتينين في الدم لجميع المجموعات التي خضعت للدراسة مع تغيرات مرضية متنوعة بما في ذلك التكتس،النخر، والنزيف، التهاب الكلية، الخراج وأخيرا الآفات الحبيبية ويعود السبب الرئيسي في هذه السمية الى الثيواستاميد.

## Introduction

Thioacetamide (TAA) is a thiono-sulfur containing compound with yellow or colorless solid crystalline. It produces sulfide ions which are used in the inorganic and organic compounds industry. Because of sulfide ions production, TAA has been used as a fungicide where it ceases the germination of fungal spores [1]. Its other applications include stabilizing motor oil, chemical reagent, textile dye, organic solvent [2, 3], leather industry, textile, paper and as a preservative of motor fuel. Thioacetamide is now used in metal salt nanoparticles production [4-6] and as a substitute for hydrogen sulfide in qualitative investigation [7, 8].

Humans can be exposed to TAA in a variety of ways [1]. It can be swallowed, inhaled or absorbed through skin. When thioacetamide is bioactivated, it produces thioacetamide S-oxide which produces peroxide radicals, subsequently causing reactive oxygen species (ROS) to develop. ROS initiates oxidation events, such as lipid peroxidation to unsaturated lipids or starting interactions with sulfhydryl molecules which results in liver damage [9, 10]. Eventually, the produced metabolites are dispersed throughout the body, including plasma, liver, bone marrow, adrenals and kidneys [11].

Exposure to TAA for longer periods causes more damage to the renal cortex than medulla due to its unequal metabolites distribution in the renal tissue where about ninety percent of the total renal blood flow arrives in the cortex via the blood stream. Therefore, relatively high concentrations of TAA metabolites can reach the cortex through blood stream before entering the medulla [12-14]. High volume of blood flow through kidney can cause damage by filtering large amount of toxins that have concentrated in the renal lobules [15]. Thioacetamide induces cells death and affects the termination of the proximal renal tubule [2]. The kidney is very susceptible to toxicants because of high blood volume that runs through it. So a large amount of toxins are filtered through it and can be concentrated in the renal tubules. It can result in systemic toxicity, thus causing a decrease in body wastes excretion ability, inability to maintain body fluid and electrolytes balance [15, 16]. Therefore, this study aimed to confirm the toxic effect of TAA in the kidneys of albino mice (BALB/c).

## Materials and Methods

### Chemicals

Thioacetamide was purchased from Qualikems, India and was administered to the mice intraperitoneally with a dose of 100 mg/kg body weight, according to [17].

### Animals

Around 3 months old one hundred twenty male albino mice with a body weight ranging 30-35g, were used to accomplish the current study. The animals were obtained from the animal house of Al-Razi Center, the Ministry of Industry, Baghdad Iraq. For adaptation mice were held in plastic cages of 60\*10\*10 cms dimensions and were kept at animal house of veterinary directorate, Ministry of Agriculture for two weeks. They were fed

on water and commercial feed pellets (standard rodent diet). Housing conditions were kept at  $22 \pm 25^\circ\text{C}$ , plus daily organized lighting using automatic electrical timer that provided a twelve hours light (07.00am- 19.00) and a twelve-hour night cycle. The cages litter was removed every 7 days.

### Study Design

Animals were randomly divided into two main groups. A control group consisted of 30 mice uptaking the normal diet only. Whereas the second treated group of ninety mice was divided into 3 subgroups as follow:

- a) 1<sup>st</sup> subgroup (G1): 30 mice treated intraperitoneally, monthly (one dose a month) for 2 months with TAA of 0.5 mg/ 100 g B.W.
- b) 2<sup>nd</sup> subgroup (G2): 30 mice treated intraperitoneally, monthly (one dose a month) for 4 months with TAA 0.5 ml/100 g B.W.
- c) 3<sup>rd</sup> subgroup (G3): 30 mice treated intraperitoneally, monthly (one dose a month) for 6 months with TAA 0.5 ml/100 g B.W.

### Blood Collection

Blood was collected for biochemical tests in different periods, 2, 4 and 6 months of experiment via cardiac puncture technique. Blood samples were collected in test tubes without using any anticoagulant and waiting for 15 minutes till clot was formed. The serum was then separated from coagulated blood samples by centrifugation at 5000 rpm for 15 minutes. Finally the serum was collected in plain tubes and was frozen at  $-20^\circ\text{C}$  until used [18].

### Biochemical Tests

Blood urea was measured by colorimetric-enzymatic method which is based on the urease action that hydrolyses urea in ammonium ions carbon dioxide complex. This coloration was measured at 600 nm which was relative to urea concentration in serum sample [17].

Serum creatinine evolution was based on alteration in the original reaction of picrate. Under alkaline state the creatinine reacted with picrate ions that lead to form a reddish complex. The concentration of creatinine on samples depended on color complex density and was determined through the increase of absorbance in a pre-fixed interval of time as described by Cannon *et al.* [19].

### Histological Evaluation

The animals were killed by chloroform and postmortem inhalation. Subsequently kidneys were macroscopically examined to document any abnormalities. Tissue specimens were collected from the organs and were fixed in 10% formalin. The specimens were later processed during routine procedure by using the histokinette [20, 21].

### Statistical Analysis

Resulting data were analyzed using statistical analysis system (SAS) by means of a complete randomized design (C.R.D.), using two way analysis of variance (ANOVA) in factorial experiment design to compare the studied parameters mean between control and the treated groups. Duncan's multiple range tests were applied [22].

### Results and Discussion

After treatment of mice with TAA, the results revealed that the mean of blood urea level in G3 ( $55.36 \pm 0.20\text{mg/dl}$ ) had significantly increased ( $P < 0.05$ ) in comparison with

concentration of G2 and G1 which were  $47.43 \pm 0.25$  mg/dl and  $40.03 \pm 0.17$ mg/dl respectively.

Regarding creatinine level in G1, G2 and G3, they were  $2.7 \pm 0.15$ ,  $3.2 \pm 0.35$  and  $4.7 \pm 0.54$  mg/dl respectively. This means there was a significant increase in the three treated groups, especially G3 group in comparison with the control.

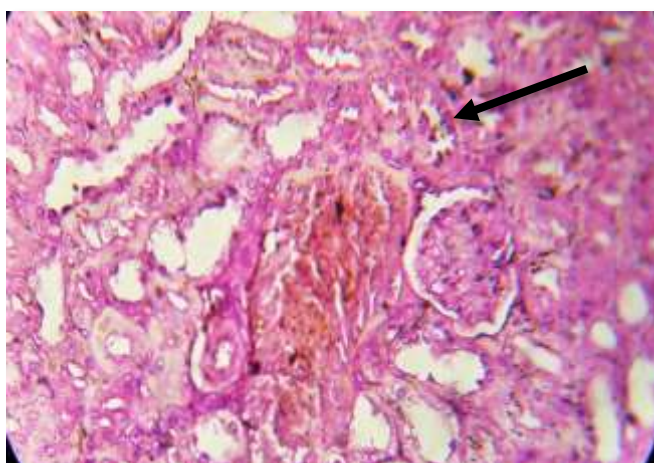
**Table 1:** The serum concentration of blood urea and creatinine of G1, G2, G3 and control show at different period

Groups	Blood Urea mg/dl	Creatinine mg/dl
Control	$23 \pm 0.20$	$0.6 \pm 0.03$
G1 2 months	$40.03 \pm 0.17^*$	$2.7 \pm 0.15^*$
Control	$23.03 \pm 0.42$	$0.60 \pm 0.05$
G2 4 months	$47.43 \pm 0.25^{**}$	$3.2 \pm 0.35^{**}$
Control	$23.05 \pm 0.25$	$0.6 \pm 0.09$
G3 6 months	$55.36 \pm 0.20^{***}$	$4.7 \pm 0.54^{***}$

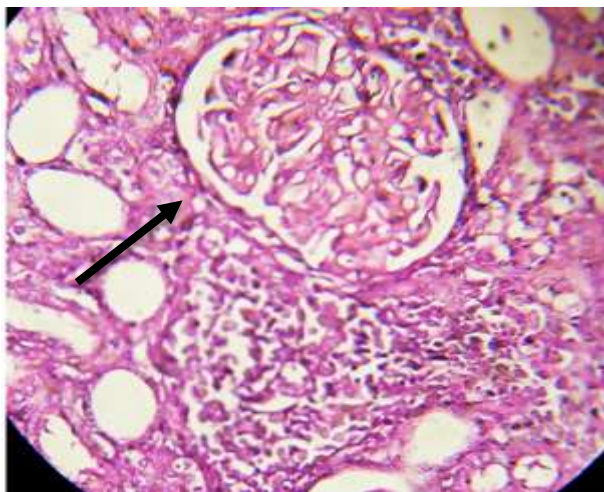
Mean  $\pm$  S.E\*significant at  $P < 0.05$ , \*\*significant at  $P < 0.01$  and \*\*\*significant at  $P < 0.001$

The histological evaluation of albino mice kidney tissue after H and E staining, showed that the kidney of control group had normal architecture. Histological changes in kidney of albino mice that were administrated with TAA I/P (100 mg/kg B.W) for the period 2 months had acute cellular swelling of renal tubules, necrosis of glomeruli and renal tubules with interstitial hemorrhage (Figure1). Whereas, after 4 months of TAA treatment, kidney showed a necrosis in renal tubules, large glomerular tuft due to an increase of mesenchymal cells and PMNCs infiltration of the interstitial (Figure 2).

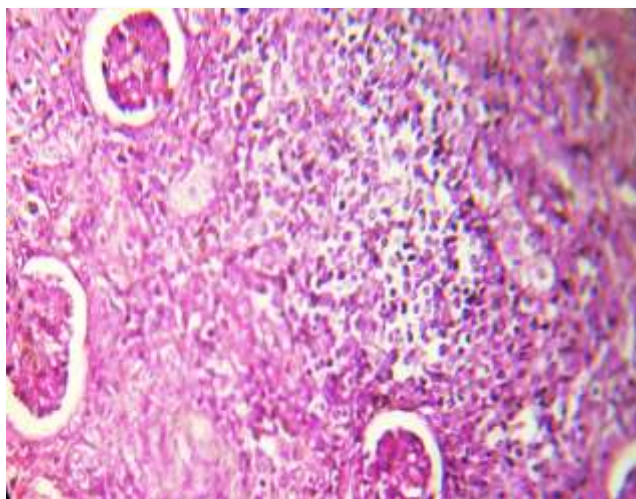
Finally, after 6 months of TAA treatment, kidneys showed granuloma with center of necrosis and MNCs infiltration that was surrounded by capsule, necrotic glomeruli, increase in the space of bowman and interstitial necrosis (Figure 3).



**Figure 1:** Kidney of albino mice administrated with TAA I/P for two months: →acute cellular swelling of renal tubules, and necrosis of glomeruli and renal tubules interstitial hemorrhage (H&E, X400).



**Figure 2:** Kidney of albino mice administrated with TAA I/P for 4 months: →necrosis of renal tubules, large glomerular tuft due to increase of mesenchymal cells and PMNCs infiltration in the interstitial (H&E, X400).



**Figure 3:** Kidney of albino mice administrated with TAA I/P for 6 months: →granuloma with center of necrosis with MNCs infiltration and surround by capsule, necrotic glomeruli, increase in the space of bowman and interstitial necrosis (H&E, X400).

The present study revealed that mice exposed to TAA exhibited histopathological changes and apparent impairment of renal function, manifested by the enhancement of urea and serum creatinine levels. Blood urea and creatinine are waste products of metabolism that are excreted by the kidney elevation of these parameters, indicating insufficiency of renal function [23]. Many previous studies have reported an increase in urea and creatinine levels in mice due to damaged kidney, causing acute renal failure [24, 25].

In addition, the toxic effect of thioacetamide has also been reported in many studies that when applied on different experimental animals, it may lead to an increase in serum creatinine levels [26-28]. This increase may occur due to a disturbance in  $\text{Na}^+\text{-K}^+$  pump of nephron and detachment of epithelial cells from renal tubules which causes nephropathy while damaging and destroying renal parenchyma [29]. Also, a study by Serag [12] reported an increase in urea

and creatinine levels in mice after getting treated with TAA. While another study by Abdou *et al.*[30] mentions that the urea level may change due to inhibition of urea cycle by inhibiting 2-oxoglutarate formation that increases ammonia which may lead to a decrease in urea concentration.

Histopathological changes, including the cortical area of kidney which was damaged more than in the medulla, were reported. The glomerular necrosis, areas of hemorrhages, cuffing, proliferation of mesangial cells and granuloma were observed. This abnormality may be due to high toxins filtration rate as a result of high blood volume flowing through kidney lobules which can concentrate in it. The renal morphology is impaired, with severe and generalized necrosis of tubular epithelial cell, diffused tubular swelling, glomerular congestion, and inflammatory cell infiltration [15].

Tubular damage and necrosis may occur as a result of decreased glomerular filtration. Tubular abnormalities may also result in tubule obstruction which causes glomerular filtrate to flow backward [1].

A study by Al-Attar *et al.*[27] mentioned that administration of TAA100mg/kg intraperitoneally for 12 weeks leads to several changes in the renal corpuscles structure, involving a high rate of necrosis and degeneration of glomeruli and Bowman's capsules due to the high volume of blood flow through them, and the filtering big amounts of toxins that can concentrate in kidney tubules. In conclusion, The TAA had a series of toxic effects on kidneys of the mice, leading to nephrotoxicity which finally leads renal failure.

## References

- [1] S. Zargar, M. Alonazi, H. Rizwana, and T. A. Wani, "Resveratrol reverses thioacetamide-induced renal assault with respect to oxidative stress, renal function, DNA damage, and cytokine release in Wistar rats," *Oxidative medicine and cellular longevity*, vol. 2019, 2019.
- [2] T. M. Chen, Y. M. Subeq, R. P. Lee, T. W. Chiou, and B. G. Hsu, "Single dose intravenous thioacetamide administration as a model of acute liver damage in rats," *International journal of experimental pathology*, vol. 89, no. 4, pp. 223-231, 2008.
- [3] J. W. Lee *et al.*, "Role of metabolism by flavin-containing monooxygenase in thioacetamide-induced immunosuppression," *Toxicology letters*, vol. 136, no. 3, pp. 163-172, 2003.
- [4] Y. Zhang, H. Wang, B. Wang, H. Yan, and M. Yoshimura, "Low-temperature hydrothermal synthesis of pure metastable  $\gamma$ -manganese sulfide (MnS) crystallites," *Journal of crystal growth*, vol. 243, no. 1, pp. 214-217, 2002.
- [5] C. Liddell and C. Summers, "Nonspherical ZnS colloidal building blocks for three-dimensional photonic crystals," *Journal of colloid and interface science*, vol. 274, no. 1, pp. 103-106, 2004.
- [6] Z. Liu, J. Liang, D. Xu, J. Lu, and Y. Qian, "A facile chemical route to semiconductor metal sulfide nanocrystal superlattices," *Chemical communications*, no. 23, pp. 2724-2725, 2004.
- [7] B. HSD, "Hazardous Substances Data Base. National Library of Medicine," ed, 2000.
- [8] I. Some anti-thyroid, "related substances, nitrofurans and industrial chemicals Monographs on the evaluation of the carcinogenic risk of chemicals to man, No 7," *Lyon: International Agency for Research on Cancer (IARC)*, pp. 245-251, 1974.
- [9] S. Zargar, T. A. Wani, A. A. Alamro, and M. A. Ganaie, "Amelioration of thioacetamide-induced liver toxicity in Wistar rats by rutin," *International journal of immunopathology and pharmacology*, vol. 30, no. 3, pp. 207-214, 2017.
- [10] S. Zargar, N. J. Siddiqi, T. H. Khan, and I. E. Elredah, "Effect of cadmium fluoride and quercetin on in vivo activity of indoleamine 2, 3-dioxygenase in mice liver and kidney," *Fluoride*, vol. 47, no. 1, pp. 31-42, 2014.
- [11] S. Ogura and T. Shimosawa, "Oxidative stress and organ damages," *Current Hypertension Reports*, vol. 16, no. 8, p. 452, 2014.

- [12] H. M. Serag, "Biochemical studies on thioacetamide toxicity in male albino rats and the role of tomato juice as an antioxidant," *Mansoura Journal of Forensic Medicine and Clinical Toxicology*, vol. 15, no. 2, pp. 99-115, 2007.
- [13] F. A. Kadir, N. M. Kassim, M. A. Abdulla, and W. A. Yehye, "Effect of oral administration of ethanolic extract of *Vitex negundo* on thioacetamide-induced nephrotoxicity in rats," *BMC complementary and alternative medicine*, vol. 13, no. 1, pp. 1-8, 2013.
- [14] E. Al-Sayed and M. M. Abdel-Daim, "Protective role of Cupressuflavone from *Cupressus macrocarpa* against carbon tetrachloride-induced hepato-and nephrotoxicity in mice," *Planta medica*, vol. 80, no. 18, pp. 1665-1671, 2014.
- [15] Q. Begum, S. Noori, and T. Mahboob, "Antioxidant effect of sodium selenite on thioacetamide-induced renal toxicity," *Pakistan Journal of Biochemistry and Molecular Biology*, vol. 44, no. 1, pp. 21-26, 2011.
- [16] T. Oduola, I. Bello, G. Adeosun, A.-W. Ademosun, G. Raheem, and G. Avwioro, "Hepatotoxicity and nephrotoxicity evaluation in Wistar albino rats exposed to *Morinda lucida* leaf extract," *North American journal of medical sciences*, vol. 2, no. 5, p. 230, 2010.
- [17] C. S. Lieber *et al.*, "Model of nonalcoholic steatohepatitis," *The American journal of clinical nutrition*, vol. 79, no. 3, pp. 502-509, 2004.
- [18] S. A. H. A. Rahman and D. A. Sattar, "Effect of different concentration of Super Cyren pesticide on some physiological and histological traits of mice after different periods of oral administration," *Iraqi Journal of Science*, pp. 2291-2300, 2017.
- [19] D. C. Cannon, R. J. Henry, and J. W. Winkelman, *Clinical Chemistry: Principles and Technics*. Medical Department, Harper and Row, 1974.
- [20] L. G. Luna, "Manual of histologic staining methods of the Armed Forces Institute of Pathology," 1968.
- [21] S. M. M. Razooki and A. M. Rabee, "Evaluation of the Toxicological Effects of Zinc Oxide Nanoparticles in Albino Male Mice," *Iraqi Journal of Science*, pp. 42-58, 2020.
- [22] A. Dmitrienko, *Analysis of clinical trials using SAS: A practical guide*. SAS Institute, 2017.
- [23] M. Y. Alomar, "Physiological and histopathological study on the influence of *Ocimum basilicum* leaves extract on thioacetamide-induced nephrotoxicity in male rats," *Saudi journal of biological sciences*, vol. 27, no. 7, pp. 1843-1849, 2020.
- [24] Y. Fazal, S. N. Fatima, S. M. Shahid, and T. Mahboob, "Nephroprotective effects of  $\beta$ -carotene on ACE gene expression, oxidative stress and antioxidant status in thioacetamide induced renal toxicity in rats," *Pakistan journal of pharmaceutical sciences*, vol. 29, no. 4, 2016.
- [25] E. Karaoz *et al.*, "Effect of chronic fluorosis on lipid peroxidation and histology of kidney tissues in first-and second-generation rats," *Biological trace element research*, vol. 102, no. 1, pp. 199-208, 2004.
- [26] M. Omnia, M. Nabila, and R. Nadia, "Biochemical effects of propolis and bee pollen in experimentally-induced hyperammonemia in rats," *Benha Vet. Med. J.*, vol. 27, no. 1, pp. 8-24, 2014.
- [27] A. M. Al-Attar, A. A. Alrobai, and D. A. Almalki, "Protective effect of olive and juniper leaves extracts on nephrotoxicity induced by thioacetamide in male mice," *Saudi journal of biological sciences*, vol. 24, no. 1, pp. 15-22, 2017.
- [28] A. Ahmad *et al.*, "Ameliorative effect of camel's milk and *Nigella Sativa* oil against thioacetamide-induced hepatorenal damage in rats," *Pharmacognosy magazine*, vol. 14, no. 53, p. 27, 2018.
- [29] E. Birkner *et al.*, "The influence of fluoride ions upon selected enzymes of protein metabolism in blood plasma of rabbits with hypercholesterolemia," *Biological trace element research*, vol. 124, no. 2, pp. 118-128, 2008.
- [30] S. E. Abdou *et al.*, "Antifibrotic Effect of Curcumin on Thioacetamide Induced Liver Fibrosis," *Alexandria Journal for Veterinary Sciences*, vol. 45, 2015.