



STUDY OF GENETIC DIVERSITY AMONG SIX IRAQI WHEAT GENOTYPES USING RANDOM AMPLIFIED POLYMORPHIC DNA

Wisam H. Salo, Ali Imad, *Gulboy A. Nasir

Institute of genetic engineering & biotechnology for post graduate studies, University of Baghdad. Baghdad-Iraq.

*College of Agriculture, Division of Basic sciences, University of Baghdad. Baghdad-Iraq.

Abstract

The degree of genetic divergence was estimated in six Iraqi soft wheat genotypes, four exotic genotypes, one produced by hybridization and one produced by gamma irradiation, through random amplified polymorphic DNA methodology. A total of 42 DNA fragments were generated by the 3 random primers, with an average of about 7.4 bands per primer. The 42 fragments showed polymorphism among the six wheat genotypes. Jaccard similarity matrix ranged from 25 to 66.7%, which indicated a high genetic diversity among the genotypes. We conclude that random amplified polymorphic DNA analysis can be used for the characterization and grouping of wheat genotypes; these results will be helpful in our wheat breeding program.

دراسة التغاير الوراثي بين ستة اصناف من الحنطة العراقية بأستعمال تقنية التضخيم العشوائي لتعدد اشكال الحامض النووي الرايبي المنقوص الاوكسجين

وسام حازم سلو، على عماد محمد منير، *كلبوي عبد المجيد ناصر معهد الهندسة الوراثية والتقنية الاحيائية للدراسات العليا، جامعة بغداد. بغداد-العراق. *شعبة العلوم الاساسية، كلية الزراعة، جامعة بغداد. بغداد-العراق.

الخلاصة

تمت دراسة الاختلاف الوراثي بين سنة أصناف من الحنطة العراقية الناعمة ، اربعة منها مستوردة وواحدة متنجة بيرامج التهجين والاخر منتج بالتشعيع، بأستعمال تقنية التضخيم العشوائي لتعدد اشكال الحامض النووي الرايبي المنقوص الاوكسجين. وتم الحصول على ٤٢ حزمة DNA بأستعمال ٣ بادئات وبمعدل ٢.٤ حزمة لكل بادئ اظهرت الحزم تغايرا بين الاصناف السعنة اذ اظهرت مصفوفة التشابه (Jaccard) نسب تشابه تراوح بين ٢٥-٦٦, % مما يؤكد وجود تغاير كبير بين الاصناف. بذلك ثبت بوضوح كفاءة تقنية التضخيم العشوائي لتعدد اشكال الحامض النووي الرايبي المنقسوم الاوكسجين في تشخيص وتصنيف الطرز الوراثية للحنطة مما يعزز برامج التهجين.

Introduction

Being a staple food, wheat occupies an important place in the crop husbandry of Iraq. But wheat production in Iraq has been decrease during recent years than it was previously, continued disimprovement in productivity conceder a highly risk because of increasing demand by the still-growing human population. However, during the last few years, yield improvement in wheat varieties has not been substantial; the narrow genetic base of the germplasm in use has been considered the main reason. Knowledge of diversity patterns allows plant breeders better to understand the evolutionary relationships among accessions, to sample germplasm in a more systematic fashion, and to develop strategies to incorporate useful diversity in their breeding programs(1). Information about genetic diversity and relatedness in the available germplasm and among elite breeding material is a fundamental element in plant breeding. The future of our breeding program depends upon the availability of genetic variability to increase productivity. Traditionally, assessment of genetic diversity has been based on differences in morphological and on agronomic traits or pedigree information for the different crops (2; 3). Recently, restriction fragment length polymorphisms

and markers isozyme have been used for diversity studies and for genetic mapping of these crop species (4; 5). But their use has remained limited, as they revealed low levels of polymorphism and isozyme expression was found to influenced by environmental be highly conditions(6). However, PCR-based DNA marker techniques seem provide to the means for generating useful genetic information on polymorphism, relatedness and diversity. The PCR-based random amplified polymorphic DNA(RAPD) markers are dominant markers and are extensively used in genetic mapping(7) and for the identification of markers linked with useful traits(8). Due to its technical simplicity and speed, RAPD methodology has been used for diversity analyses in crops several (9). Wheat is large characterized by а genome size (approximately 17,000 Mb) and little or no information sequence is available for the wheat genome. We made an RAPD analysis of six genotypes of wheat to genetic estimate their diversity and relatedness and to compare the different source genotypes. The information gathered will be helpful for our breeding programs.

Material and Methods

The plant material: The plant material used in the study consisted of six genotypes of wheat:

Table 1:	The wheat	var	ieties that	used	in study	
¥7	N	. C	0			

Variety	Method of production	Origin
Al-nedaa	γ-rad 15Kr	Saber beak X maxibac
Al- tahady	Hybridization	Saber beak X maxibac
Al-noor	Exotic	Ecarda
Al- hashimia	Exotic	Ecarda
Um rabee'a	Exotic	Jori69 X Haw
Wahat al-Iraq	Exotic	Pic "S"Roffs X Rhexta

All genotypes were planted in pots in a growth chamber.

DNA extraction:

The wheat genotypes were grown in plastic containers (250 mL), 0.2-0.3 g leaf tissues were obtained from the 6-day-old seedlings. 100mg of newly grown leaf used to extract DNA by CTAB lyses as listed in (10).

RAPD analyses:

DNA concentrations in the working solution of approximately $15ng/\mu L$ in d.H2O were confirmed by spectrophotometer. For RAPD analysis (11), 1X PCR buffer, 200 μ M of each dNTPs, 10 pMol random primers and 1.25 U of *Taq* DNA polymerase, concentration of genomic DNA and MgCl₂ were optimized. The 10-base oligonucleotide primers obtained from alfa DNA Canada as listed in table 2. DNA amplification reactions were performed in a thermal cycler (GeneAmp 9700, ABI). The PCR profile was: one cycle of 94°C for 5 min, 40 cycles of 94°C for 1 min, 38°C for 1 min, and 72°C for 1 min, and a final extension for 10 min at 72°C.

Table 2: The primers that used in study			
OPA 05	5'-AGGGGTCTTG-3'		
OPB-05	5'-CCTTCACGCA-3'		
OPC-05	5'-GATGACCGCC-3'		

Analyses of RAPD data:

The RAPD fragments were analyzed by electrophoresis on 1.5% agarose gels with ethidium bromide (10 ng/100 mL of agarose solution in Tris borate EDTA buffer). The bands were counted by starting from the top of the lanes to the bottom. All visible and unambiguously scorable fragments amplified by the primers were scored under the heading of total scorable fragments. Amplification profiles of the six genotypes were compared with each other, and bands of DNA fragments were scored as present or absent.

The data of the primers were used to estimate genetic similarity (Table1) on the basis of number of shared amplification products (12). The equation used was: No. of shared amplification products = $2 \times (No. of common bands between any two lanes) / (Total No. of bands in the same two lanes). Genetic relationship among the genotypes was estimated with the dendrogram (Figure 1) constructed using unweighted pair group of arithmetic means UPGMA (13).$

Results and Discussion

DNA of six varieties of wheat was amplified with 3 different random primers, as listed in table 2, fragments were generated by the 3 primers, with an average of about 7.4 bands per primer (range = 3-11). The number and size of the DNA fragments were strictly dependent upon the sequence of the primer. Reactions were repeated from two to three times to check the consistency of the amplified products; only easily resolved and bright DNA bands were counted. All the genotypes differed, based on their amplification profile with these 42 DNA bands. 32 fragments were polymorphic (76%) in these six wheat varieties. Ten individual plants of each genotype were tested separately; all showed similar banding patterns, indicating that the genotypes were highly homozygous. All the six wheat varieties could be identified with a single primer. These results suggest that RAPD markers provided substantial information for the identification of wheat genotypes. Among the six wheat genotypes, Al-tahady produced the largest number of DNA-amplified fragments (12 bands), while the smallest number (4 bands) was produced by Al-noor.

The reproducibility of the RAPD technique can be influenced by variable factors, such as primer sequence, template quality and quantity, the type of thermocycler, and polymerase concentration (7). However, the use of a standardized RAPD protocol can ensure a reproducible RAPD pattern.

The concentrations of MgCl₂, *Taq* DNA polymerase and concentrations of template DNA were optimized for PCR conditions. DNA concentrations of 5, 10, 15, 20, and 25 ng/25 μ L in each reaction were assayed. The concentration of 10 ng/25 μ L was found to

produce the most consistent and reproducible banding patterns. Tahir(14) used RAPD technique to evaluate Iraqi wheat mutants and found that 3mM MgCl₂ was the optimum concentration for amplification. same results have been obtained. Higher than 3mM MgCl₂ produced nonspecific amplification. Similarly, one unit concentration of *Taq* DNA polymerase was found optimum for amplification of genomic DNA. Other reaction conditions were also kept constant, and the results were found to be consistent and reproducible. All of the amplified bands were identical in each repetition.



Figure 1: An example of RAPD banding pattern obtained from primer OPB-05 on 15% agarose gel, 5V/cm at 1hr. for 6 genotypes of wheat, a1: Al-nedaa, a2: Altahady, a3: Al-noor, a4: Al-hashimia, a5: Um rabee'a, a6: Wahat al-Iraq and lane M represented the molecular marker (100bp DNA Ladder Promega).

Table 3: Similarity Matrices computed with Jaccard coefficient for 6 wheat varieties* obtained from RAPD-PCR showing the relationship

	8
between	verities.

Simil	larity icient	Matrix	Matrix computed with Jaccard			
totii	a1	a2	a3	a4	a5	ab
al	1	0.643	0.16 7	0.250	0.364	0.364
a2		1	0.214	0.286	0.286	0.286
a3			1	0.500	0.286	0.500
a4				1	0.66 7	0.250
a5					1	0.250
aб						1

*a1: Al-nedaa, a2: Al- tahady, a3: Al-noor, a4: Alhashimia, a5: Um rabee'a, a6: Wahat al-Iraq



Figure 2: Dendrogram of six wheat varieties showing genetic similarity based on RAPD data by using UPGMA cluster analysis, showing the relationship between verities.

The line Al-Nedaa was found to be 64.3% similar to Al- Tahady. Both these genotypes will be has the same breeding program, the difference between them is due to the gamma

irradiation which may cause changes in there genomics. Similarly, Al-Noor was 50% similar to line Wahat al-Iraq, as it is seen in table (1) exotic both lines is and may share the same origin. Al-Hashimia and Um rabee'a found to hase 28.6% similarity. The last four genotypes clustered in second group. Al-Nedaa and Al- Tahady are placed in first group. This clustering of the genotypes might be due to the selection from a single population: the same with was observed other wheat cultivars (15). The low similarity of Al-Nedaa and Al- Tahady with the rest of the genotypes seems to be due to the fact that they are only local varietis. Bibi et al. (16) compared hemp varieties using RAPDs. They found a mean of 97.1% polymorphism over all varieties and loci. Mukhtar et al. (15) observed 445 DNA amplified fragments with 50 random primers in 20 varieties; they found 64.38% polymorphism. In our study, 42 DNA fragments were amplified with 3 random primers, with an average of 7.4 bands /primer. There was 45% polymorphism in the six wheat varieties. Overall, a high genetic base was found, with 16.7 to 64.3% similarity among the six genotypes. RAPD technique was found to be quite effective in determining the genetic variation among wheat genotypes and could be utilized as DNA fingerprinting for variety identification and for the establishment of plant breeder rights in Iraq. These findings would also contribute to choose parents for our breeding program.

References

1. Tandon, P.; Kumaria, S. and Nongrum, J. **2009**. Conservation and management of plant genetic resources in north India.Indian Journal of Traditional Knowledge, 8: 29-34.

- 2. Ojaghi, J. and Akhundova, E. **2010** Genetic diversity in doubled haploids wheat based on morphological traits, gliadin protein patterns and RAPD markers. African Journal of Agricultural Research, 5: 1701-1712.
- 3. Sawalha, K.; Eideh, H.; Laham, S.; Hassan, H.and Mezeid, B.**2008** Genetic Diversity Studies on Wheat Landraces in Palestine Using RAPD Markers in Comparison to Phenotypic Classification. Journal of Applied Biological Sciences, 2: 29-34.
- Chao, S.; Sharp, A.J.; Worland, E.J. Warham, R.M.D. Koebner, and M.D. Gale. **1989**. RFLP-based genetic linkage maps of wheat homoeologous group 7 chromosomes. Theoretical Appied Geneics. 78:495-504.
- Paull, J.G; Chalmers K.J; Karakousis A. Kretschmer, J.M, **1998**. Genetic diversity in Australian wheat varieties and breeding material based on RFLP data. Theoretical Appied Geneics.96: 435-446.
- 6. Siddiqui, M. F. and Naz, N.**2009** Protein landmarks for diversity assessment in wheat genotypes. African Journal of Biotechnology. 8: 1855-1859
- Siddiqui, M.; Iqbal, S.; Erum, S.; Naz, N. and Khan, S.2010 DNA Landmarks for genetic relatedness and diversity assessment in Pakistani wheat genotype using RAPD markers. Pakistani Journal of Botany. 42: 1013-1020.
- 8. Bai, G.; Guo, P. and Kolb, F.L. **2003**. Genetic relationships among head blight resistant cultivars of wheat assessed on the basis of molecular markers. Crop Science. 43: 498-507.
- Priolli, R.; Pinheir, J. Zucchi, M.; Bajay, M. and Vello, N. 2010. Genetic Diversity among Brazilian Soybean Cultivars Based on SSR Loci and Pedigree Data. Brazilian Archives of Biology and Technology.53: 519-531

- 10. Doyle, JJ.; Doyle, J. L. **1990**. Isolation of plant DNA from fresh tissue. Focus. 12: 13–15.
- Williams, J.G.K.; Kubelik, A.R.; Livak, K.J.; Rafalski, J.A. and Tingey, S.V. **1990**. DNA polymorphisms amplified by arbitrary primers are useful as genetic markers. Nucleic Acids Research. 18: 6531-6535.
- 12. Jaccard, P. **1908**. Nouvelles recherches sur la distribution florale; Bulletin Societe Vaudoies Sciences Naturelles, 44: 223–270.
- 13. Sneath, P. and Sokal, R. **1973**. Numerical Taxonomy. Freeman, San Francisco.32-171.
- 14. Tahir N. A. 2008. Assessment of Genetic Diversity among Wheat Varieties in Sulaimanyah using Random Amplified Polymorphic DNA (RAPD) Analysis. Jordan Journal of Biological Sience.1:159-164
- Mukhtar, MS., Rehman, M; and Zafar, Y. 2002. Assessment of genetic diversity among wheat (*Triticum aestivum* L.) cultivars from a range of locations across Pakistan using random amplified polymorphic DNA (RAPD) analysis. Euphytica. 128: 417-425.
- 16. Bibi, S.; Dahot, M.; Khan, I.; Khatri, A. and Naqvim. 2009. Study of genetic diversity in wheat (Triticum Aestivum L) using random amplified polymorphic DNA (RAPD) markers. Pakistani Journal of Botany.41: 1023-1027