



Beauveria bassiana (Bals.) Vuill.

*

*

) *Beauveria bassiana* (Bals.) Vill
1± 28 4 3 2 1 ()
(/ 8)
(P<0.05)
/ 10⁵×68 69 67
(P<0.05)
%24.3
(P<0.05)
50/ 0.507
/ 10⁵× 30 19
(50/ 0.2)

THE EFFECT OF SOME CARBON SOURCES ON GROWTH PARAMETERS OF THE *BEAUVRIA BASSIANA* (BALS.)VUILL

Hussein Magtoff Diwan, *Muna Hemudi Khamas

Department of Biological Control of Plant Pathogens, Center of Integrated Control, Directorate of Agricultural Research, Ministry of Science & Technology. Baghdad - Iraq.

*Department of Biology, College of Science, University of Baghdad. Baghdad - Iraq.

Abstract

Seven carbon sources; Glucose, Sucrose, Soluble starch, Pectine, Dibs, Wheat seeds extract and Rice grains extract individually incorporated in different liquid media were evaluated for their effects on sporulation, percent of spores germination and fungal biomass dry weight of entomopathogenic fungus *Beauveria bassiana* for four incubation periods ;1,2,3 and 4 days at 28±1c. Carbon used in the basal medium as 8g/l. The results showed that glucose, rice grains extract, wheat seeds extract were significantly superior (p<0.05) to induce the fungal sporulation which recorded 67,69,68×10⁵ spore/ml after 4 days of incubation, respectively. Soluble starch was

more efficiency for spores germination, whereas the percent of spores germination was 24.3 after 4 days of incubation. Rice grains extract increased significantly the dry weight of fungus which recorded 0.507g/ml after 4 days of incubation. Pectine, soluble starch, sodium nitrate, methionin and glycine were less efficiency for biomass production (0.2g/50ml).

(10 9)
)
 (Corn flour
 . (2 1)
 . (10 11)
 (3)
 . (4)
 (5) . (14 13 12) *B. bassiana* (5)
 Boverin
 Sterile (IPC-P) *Paecilomyces lilacinus* *B. bassiana* . (6) Mycotrol
 (ARF907) hyphum
 glycines *Meliodyne incognita*
Heterodera ()
Trichoderma viride
Pochnia
 (HSY-12-14) *chlamydosporia* (7) *B. bassiana*
 . (8)
 (9)
)
 () ()
 .*B. bassiana*
Beauveria bassiana (Bals.) Vuill IBC) *B. bassiana*
 . (5) (1201)
 B. Specific spores
 (9)
bassiana *B. bassiana*
 (PSA)

-

/

(10)

haemocytometer

AB

B. bassiana

(17)

SPSS

(p=0.05)

Beauveria bassiana

(1)

(P < 0.05)

B. bassiana

/ $10^5 \times 17$

/ $10^5 \times 33$ 36

B. bassiana

/ $10^5 \times 43$

$\times 15$ / $10^5 \times 11$ / $10^5 \times 21$

/ 10^5

(P < 0.05)

(/ $10^5 \times 68$ 69 67)

/ $10^5 \times 50$ 19 30 50

1±28

4 3 2 1

B. bassiana

()

10 (10) PDA

(%0.05) 20

(5)

(/ $10^6 \times 2.3$)

: Haemocytometer

0.2) = /

$\times 10^4 \times 25 \times (0.2 \times$

(0.5) MgSO4 (0.5) K2HPO4 (1)

100) FeSO4 (0.01) KCl

(ppm)

(22.2) (10) (20)

(28.6) (22.6)

(36.4) ()

(16 15)

pH 6.5

1N NaOH

.1N HCl

:***Beauveria bassiana***

/ 50

جدول ١: تأثير بعض المصادر الكربونية في القابلية على التبوغ والنسبة المئوية لإنبات الابواغ والوزن الجاف للقطر *B. bassiana* بعد مدد حضن مختلفة عند درجة حرارة 28±1 م°

المصدر الكربوني	معدّل النمو	١ يوم	٢ يوم	٣ يوم	٤ يوم
كلوكوز	انتاج الابواغ × 10 ^٥ بوغ/م ^٢	C 16.7 ^a ± 4.40	B 35.8 ^a ± 2.20	B 43.3 ^{ab} ± 1.66	A 66.7 ^a ± 7.26
	نسبة الأنبك %	A 3.3 ^a ± 3.3	A 15.4 ^a ± 7.98	A 15.3 ^a ± 1.4	A 4.4 ^a ± 2.73
	الوزن الجاف غ/50م ^٢	C 0.233 ^a ± 0.00	B 0.350 ^a ± 0.00	B 0.387 ^a ± 0.00	A 0.443 ^b ± 0.02
سكروز	انتاج الابواغ × 10 ^٥ بوغ/م ^٢	C 16.7 ^a ± 4.16	B 32.5 ^a ± 3.81	B 36.7 ^b ± 3.33	A 50 ^b ± 1.44
	نسبة الأنبك %	A 6.7 ^a ± 6.66	A 10.5 ^a ± 6.18	A 20.4 ^a ± 3.39	A 8.3 ^b ± 1.49
	الوزن الجاف غ/50م ^٢	D 0.100 ^b ± 0.01	C 0.220 ^b ± 0.00	B 0.313 ^b ± 0.04	A 0.393 ^b ± 0.00
نشا	انتاج الابواغ × 10 ^٥ بوغ/م ^٢	C 1.9 ^b ± 0.22	B 18.3 ^{bc} ± 0.06	B 21.4 ^c ± 2.03	A 29.7 ^c ± 2.95
	نسبة الأنبك %	A 21.3 ^a ± 2.43	B 2.7 ^a ± 1.55	15.6 ^a A ± 7.2 B	A 24.3 ^a ± 3.73
	الوزن الجاف غ/50م ^٢	D 0.043 ^c ± 0.00	C 0.123 ^c ± 0.00	B 0.190 ^c ± 0.00	A 0.200 ^d ± 0.00
بكتين	انتاج الابواغ × 10 ^٥ بوغ/م ^٢	C 2.2 ^b ± 0.57	B 9.5 ^c ± 0.14	B 10.9 ^c ± 0.37	A 19.4 ^c ± 3.13
	نسبة الأنبك %	A 6.9 ^a ± 6.9	A 4.4 ^a ± 3.15	A 4.4 ^a ± 2.21	A 3.3 ^a ± 2.14
	الوزن الجاف غ/50م ^٢	C 0.193 ^a ± 0.04	B 0.300 ^a ± 0.01	A 0.363 ^a ± 0.00	A 0.423 ^b ± 0.01
دبس	انتاج الابواغ × 10 ^٥ بوغ/م ^٢	B 3.1 ^b ± 2.9	B 11.6 ^c ± 0.52	B 14.9 ^c ± 1.36	A 44.9 ^b ± 5.03
	نسبة الأنبك %	A 16.0 ^a ± 8.30	A 4.2 ^a ± 2.72	A 28.5 ^a ± 15.06	A 12.5 ^b ± 2.7
	الوزن الجاف غ/50م ^٢	B 0.240 ^a ± 0.00	B 0.283 ^a ± 0.00	A 0.403 ^a ± 0.02	A 0.370 ^a ± 0.02
مستخلص حبوب الرز	انتاج الابواغ × 10 ^٥ بوغ/م ^٢	D 3.0 ^b ± 0.57	C 23.7 ^b ± 2.68	B 55.8 ^a ± 6.00	A 69.1 ^a ± 2.52
	نسبة الأنبك %	A 16.0 ^a ± 1.85	B 1.7 ^a ± 1.66	AB 8.1 ^a ± 3.86	AB 9.7 ^b ± 2.46
	الوزن الجاف غ/50م ^٢	D 0.080 ^b ± 0.00	C 0.203 ^b ± 0.01	B 0.410 ^a ± 0.03	A 0.507 ^a ± 0.02
مستخلص بذور الحنطة	انتاج الابواغ × 10 ^٥ بوغ/م ^٢	C 1.7 ^b ± 0.16	C 13.3 ^a ± 1.49	B 50.8 ^a ± 8.45	A 67.5 ^a ± 2.50
	نسبة الأنبك %	A 11.1 ^a ± 11.1	A 19.6 ^a ± 7.1	A 7.7 ^a ± 1.72	A 9.8 ^b ± 0.9
	الوزن الجاف غ/50م ^٢	D 0.040 ^c ± 0.00	C 0.123 ^c ± 0.00	B 0.363 ^a ± 0.03	A 0.447 ^b ± 0.00

*

) (
 (15) P<0.05
 × 56 (
 / 10⁵ × 51 / 10⁵
 (16)
 46) (/ 42.5) 10⁵×43
 (/
 (26) *B . bassiana*

B . Nomuraea rileyi *Metarhizium anisopliae* global spores
bassiana
 10⁸ × 1.1 ± 0.01 10⁸ × 0.01 ± 0.03
 / 10⁸ × 4.3 ± 0.1 / (9)
 (3)
 ()
B. brongniartii *B. bassiana* *B . bassiana*
 10⁹ × 4.38 12 (%100 – 30)
 / 10⁹ × 3 /
 6
) (18)
 (

Fusarium SP Macroconidia
B . bassiana

B . bassiana

(5)

B . bassiana

(1)

/ 10⁷ × 29.4 15.1

B. bassiana

B . bassiana

(%21.3)

3 2

%50 %55)

%11.1 %16.6

%15.6 %2.7

.%24.3

B. bassiana

%19.6

(%28.5)

()

(9)

B. bassiana

Bassianolide

microcycle

(24)

(22 21 20 7)

conidiation

.(1)

%

B .

bassiana

(1)

B. bassiana

(23)

4 2

(P < 0.05)

%4.4 %3.3

0.387 0.350 0.233)

) (50 / 0.443

/ 0.423 0.363 0.300 0.193

4 1

4 3 2 1 (50

%8.3 %6.7

(P < 0.05)

3 1

%4.2

50 / 0.403 0.240

(P < 0.05)

%21.3

3

4

B . bassiana
Trichoderma spp.

50 / 0.447 0.363

50 / 0.507 0.41

3 1

B . brongniartii *B . bassiana*

(3)

Reference (المصادر)

1. Lewis, L.C. and J.E. Cossentine .(1986)

Season long intraplant epizootics of entomopathogens, *Beauveria bassiana* and *Nosema pyrausta* in a corn agroecosystem. Entomophaga 31:364-369.

(P < 0.05)

(50/ 0.507)

2. Burges, H. D. (1998). Formulation of Microbial Biopsticides. Kluwer Academic publishers. London, U.K. 412pp

(50/ 0.447)

(1) (50/ 0.443)

3. Nelson, T. L. , A. low and T. R. Glare. (1996) .Large scale production of New Zealand strains of *Beauveria* and *Metarhizium* .In: Proceeding of the 49th New Zealand Plant Protection Conference; 1996 Aug.13-15;Nelson,New Zealand Plant Protection Society;P.257-261.

85 – 65

(25) %30 – 16 %

2 1

3

4. Rombach,M.C.,R.M. Aguda and D.W. Roberts. (1988).Production of *Beauveria bassiana* in different liquid media and subsequent conidiation of dry mycelium.Entomophaga.33:315-324.

mannan xylans Galactose

(24) Arabinose

P < 0.05

5. Sun, M. H. and X. Z. Liu. (2006) . Carbon requirements of some nematophagous entomopathogenic and mycoparasitic Hyphomycetes as fungal biocontrol agents . Myopathological. 161 : 295 – 305 .

4 3

6. Wan, H. (2003) . Molecular biology of the entomopathogenic fungus *Beauveria bassiana*: Insect-cuticle degrading enzymes and development of a new selection marker for fungal transformation. A thesis submitted to University of Heidelberg in partial fulfillment of the requirements for degree of Doctor of philosophy in Natural Sciences. 165pp.

B . bassiana

7. Thomas,K.C.,G. Khachatourians and W. M. Ingledew, (1987).Production and properties of *Beauveria bassiana* conidia cultivated in submerged culture. Can.J.Microbiol. 33: 12-20.

P. M. anisopliae *B . bassiana*
fumosoroseus

(26)

18. Bilgrami, K. S. and R. N. Verma. (1988). Physiology of fungi. Translated by Sarhan, A.R.T. and sharif, F.M. Saladin University High Education and Scientific research Ministry.Mosul University Pulpication-Mosul. 596pp.
19. Deshpande, M.V.and, U. Tuor . (2001). Microbial control of Pests: Entomopathogenic Fungi as Mycoinsecticides. Eidgenossische Technische Hochschule ETH,Institut für Mikrobiologie, Switzerland. (Report).
20. Feng,M.G.;T.J.PoprawskiandG.G.Khachatourians.(1994).Production,formulation and application of the entomopathogenic fungus *Beauveria bassiana* for insect control :Current status .Biocont.Sci.Technol.4:3-34.
21. Jaronski,S.T.andM.S.Goettel.(1997).Development of *Beauveria bassiana* for control of grasshoppers and locusts.Mem. Entomol. Soc. Can. 171:225-237.
22. Jenkins, N.E. and C. Prior. (1993). Growth and formation of true conidia by *Metarhizium flavoviride* in a simple liquid medium . Mycol. Res . 97: 1489-1494.
23. Müller,E. and W. Loeffler. (1976).Mycology.Translated by Bryce Kendrick and Barlocher.Georg Thieme Publishers Stuttgart.306pp.
24. Samsinakova,A.(1966).Growth and sporulation of submersed cultures of the fungus *Beauveria bassiana* in various media.J Invertb.Pathol.8:395-400
25. Perry, D.F. and J.P. Latge. (1980) Chemically defined media for growth and sporulation of *Entomophthora Virulenta*. J.Invertb. Pathol . 35: 43-48.
26. Vaughan, L. and H. Warren. (2000). steam – exploded agricultural wastes as a novel source of nutrients for production of *Metarhizium anisopliae*. Abstracts of 33th Ann. Meeting Guanajuato, Mexico . University of Guanajuato. PP.114
8. Stanbury, P. F., A. Whitaker and S. J. Hall. (1995). Media for industrial fermentations. In principles of fermentation technology Edited by P. F. Stanbury, A. Whitaker, and S. J. Hall. Elsevier science Ltd, New York, Tokyo pp. 93-122.
9. Bidochka, M. J., T. A. Pfeifer and G. G. khachatourians. (1987). Development of the entomopathogenic fungus *Beauveria bassiana* in liquid cultures. Mycopathologia 99: 77-83.
10. Lacey, A. L. (1997). Manual of techniques in insect pathology. Academic press, New yourk 410 pp.
11. Boyette, C. D., P. C. J. Quimby, W. J. Connick, D. J. Daigle and F. E. Fulgham.(1991). Progress in the production, formulation, and application of mycoherbicides. In Microbial control of weeds. Edited by D.O. Te Beest. Capman and hall, New York, pp.209-222.
12. Parker, B. L., M. Skinner, M. Brownbridge and M. EL-Bouhssini (2000). Control of insect pests with entomopathogenic fungi. Arab. J.pl. prot. 18: 133-138.
13. Jenkins, N. E., G. Heviefo, J. Langewald , A. J. Cherry and, C .J. lower . (1998). Development of mass production technology for aerial conidia for use as mycopesticides. Biocontrol New and Informa .19:21N-31N.
14. Posada-Florez. J. (2008) . Production of *Beauveria bassiana* fungal spores on rice to control the coffee berry borer, *Hypothenemus hampei* , in Colombia . J. Ins . Sci.8: 1536 -2442.
15. Dubios, N., K.A. Gilles, J.K. Hamilton, P.A. Rebers and F.Smith. (1956). Colorimetric method for detection of sugars and related substances. Annal. Chem .28: 350-356.
16. Lowry, O.H., NJ Rosbrough, Al- Farr, and RJ Randall. (1951). Protein measurement with the folin phenol reagent. J. Biol. Chem. 193: 265-275.
17. .Diwan,H.M.(2010).Preparation of bio-formula of *Beauveria bassiana*(Bols.) Vuill.and evaluate its efficiency against the cabbage aphid *Brevicoryne brassicae* (L.).A thesis submitted to college of science,University of Bagdad in partial fulfillment of the requirements for degree of Doctor of philosophy in Biology / Botany.165pp.