Jaddoa et al.

Iraqi Journal of Science, 2024, Vol. 65, No. 10, pp: 5499-5505 DOI: 10.24996/ijs.2024.65.10.15





ISSN: 0067-2904

Effectiveness of *Eucalyptus camaldulensis* Leaves Oil in Upregulating *exoU* expression in *Pseudomonas aeruginosa*.

Nihad T. M. Jaddoa¹, Mahmood A. H. AL-Sheikhly², Israa A.M. Aldobaissi¹, and Harith J. F. Al-Mathkhurv¹

¹Department of Biology, College of Science, University of Baghdad, Baghdad, Iraq, 10071 ²Laboratory Department, Directorate of Technical matters, Ministry of Health, Baghdad, Iraq

Received: 21/5/2023 Published: 30/10/2024 Accepted: 2/9/2023

ABSTRACT

Results of the current study demonstratedthat out of eighty-three isolatesof Pseudomonas aeruginosa, only twenty-five isolates were resistant to five different antibiotics (of different classes) that were consequently considered multidrug resistant isolates.These isolates developed variable susceptibility toward Eucalyptuscamaldulensisleavesoil (ECO). GC-MS analysis of ECOrevealed that the aromatic oil eugenol is the major constituent. However, the most frequent MIC was 0.39 µg/ml, while the lowest frequent MIC was 3.125 µg/ml.Moreover, this oil at $\frac{1}{2}$ MIC (0.195 μ g/ml) increased the gene expression of *exoU*. It is concluded from the outcomes of the studythat ECOmay cause severe damagewhen used to treat infections caused by P. aeruginosa.

Keywords: Eucalyptuscamaldulensis, Pseudomonas aeruginosa, exoU, Leaves oil

فعالية زيت اوراق Eucalyptus camaldulensis في زيادة التعبير الجيني لجين exoU ىكترىاPseudomonas aeruginosa

 1 نهاد طه محمد جدوع 1 ،محمود عبد الرزاق الشيخلى 2 ، اسراء عبد الرزاق الدبيسى 1 ، حارث جبار المذخورى ¹ قسم علوم الحياة، كلية العلوم، جامعة بغداد، بغداد، العراق ² وزارة الصحة، دائرة الأمور الفنية، قسم المختيرات، مدير الشعبة الحكومية

الخلاصة

أظهرت نتائج الدراسة الحالية انه من مجموع ثلاثة وثمانون عزلة من بكتريا الزائفة الزنجارية، خمسة وعشرون عزلة فقط قاومت خمسة أنواع مختلفة من المضادات الحياتية والتى اعتبرت عزلات متعددة المقاومة وقد تم اختبار هذه العزلات تحت تاثير زيت أوراق للمضادات اليوكالبتوس (ECO). أظهرت النتائج حساسيه متغيره تجاه الزيت المستعمل ، ويشفت تحليلات GC-MS لتحكان المكون الرئيسي للزيت العطري هو االيوجينول. واظهرت نتائج التركيز المثبط الادنى ان قيمة التركيز المثبط الادنى الأكثر تكرارا بلغت 0.39 ميكروغرام / مل. بينما كانت قيمة التركيز المثبط الادنى الاقل تكرارا 3.125ميكروغرام / مل. علاوة على ذلك، فإن هذاالزيت عند التركيز تحت المثبط الادنى (0.195 ميكروغرام / مل)قد زاد من التعبير الجينى لجين exoU .يستنتج من خلال هذه الدراسة، ان زيت اليوكالبتوس قد يتسبب في اضرار شديدة عند استعماله لعلاج الاخماج الناجمة عن بكتريا P. aeruginosa.

1. Introduction

Pseudomonas aeruginosa, an aerobic, motile, gram-negative rod like bacterial isolate, is widely distributed in varioushabitats[1]. Moreover, since multidrug-resistant *P. aeruginosa* is also emphasised as a significant concern by the World Health Organization due to it beingcarbapenem-resistant, priority of the highest level for the need to develop novel antibioticsalso requires great attention[2].

This opportunistic pathogen can cause numerous acute and chronic infections which can result in high mortality rates in a variety of hosts and organs due to its extensive repertoire of virulence factors and complex regulatory network of intra- and inter-cellular signals enabling it to adapt, flourish, and evade host defences. A key predictor of virulence has been discovered as the Type-III secretion systems (T3SS) which is responsible for injecting*ExoS*, *ExoT*, *ExoU*, and *ExoY*exotoxins inside the host cell. Nevertheless, ExoU is indicated as the most toxic as well as being the most important exotoxin that is related to epithelial injury in variousinfections. The release of ExoU leads to the fast destruction of the host cell cytoplasmic membrane [3].

Remarkably, there is a global apprehension that the extensive use of antibioticshas resulted in the establishment and dissemination of organisms with multiple antibiotic resistance that has further complicated the therapeutic strategies. Such worrisome discovery has expaneds the horizons for using medicinal plants as natural sources for new antimicrobial agents [4]. Many people all around the world have used plants for medicinal purposes to cure or prevent infections, although large number of these herbal medications have not been tested for their safety and efficiency. Additionally, the purity and potency of such herbal products are not controlled, therefore, most of inacceptable effects might be brought on by contamination or batch-to-batch variations. Meanwhile, the potential for several negative consequences may be increased by the potency of such herbal products [5].

Of interest, *E. camaldulensis*a well-known plantin Australia, belongs totheMyrtaceae family and is commonly known as river red gum[6].Many medicinal benefits have been implicated in*E. camaldulensis*leaves.The phytochemical investigationhas revealed the presence of severaldifferent secondary metabolites with antimicrobial activityin *E. camaldulensis*leaves oil [7].

Taking that into consideration, the present study aimed to inspect theimpact of ECO on the gene expression of *exoU* in *P. aeruginosa*.

2.Materials and Methods

1. Plant Description and Phytochemical Analysis

- Plant Description

The main macromorphological properties were described to simplify the identification of studied tree as proposed by Al- Masoudi & Al-Dobaissi.[8].

An amount of 100 g of dried *E. camaldulensis* leaves were crushed and then hydro distilled for 4 hr. in Clevenger apparatus to extract ECO which was later stored in glass vials at $4-5^{\circ}$ C until required [9, 10].

- Gas Chromatograph-Mass Spectrometry (GC-MS)Analysis

An AOC-20i auto-injector-equipped GC-MS Shimadzu QP 2010 plus was used to analyse the oil from *E. camaldulensis* leaves (Shimadzu, Japan). The GC temperature was set to be 80°C for the column, 280°C for the injection, helium as the carrier gas, split injection mode, and pressure flow control mode. The column flow was 1.46 ml/min with total flow being 11.8

mL/min, the linear velocity was 44.5 cm/sec, the purge flow was 3.0 mL/min, the split ratio was 5.0, and 100.0 kPa pressure. GCMS-QP 2010 plus program was used and the conditions were: Detector Gain of 0.70 kV, interface temperature of 280°C, and ion source temperature of 200 °C. Scan speed was increased from 50 m/z to 800 m/z [11]. By matching the spectra data with a known molecule that was stored in the NIST library, the sample components were identified [12] which were confirmed according to peak area, retention time and molecular formula.

2. Pseudomonas aeruginosa Isolates

A total of 83 isolates of *P. aeruginosa* were obtained from the Microbiology lab at the Department of Biology, College of Science, University of Baghdad. These isolates were originally isolated from different specimens including blood, wound swabs, sputum, and mid-stream urine. All these isolates were previously identified by amplifying 16SrRNA.

3. Antimicrobial Susceptibility Test

The Kirby-Bauer method was performed to detect the antibiotic susceptibility of *P*. *aeruginosa* isolates towards five distinct therapeutically significant antibiotics: Ciprofloxacin (5µg/disk), amikacin(30µg/disk), gentamicin (10µg/disk), imipenem(10µg/disk) and ceftazidime(30µg/disk).Clinical and Laboratory Standards Institute guidelines (CLSI) were applied to categorize an isolate if it was sensitive, intermediate and resistant [13].

4. Assay for Minimal Inhibitory Concentration

CLSIapproved microtiter broth dilution method was used to determine the minimal inhibitory concentration (MIC) of ECO [13]. The MIC is the lowest ECO concentration at which there was no visiblegrowth was detected. The ECO was serially double-diluted in Mueller-Hinton broth from 12.5 to $0.024\mu g/ml$. which were prepared in a 96-well plate (100 μ l per well). Wells without ECO wereregarded as positive growth control. A final concentration of 5×10^5 CFU/ml was achieved by adding a diluted bacterial sample to each well which was verified by viable counts.Bacteria-freewellsserved as negative growth control. The plates were kept in the incubator for 24 hours at 37°C [14].

5. Gene Expression

The effects of ECO at $\frac{1}{2}$ MIC on the expression levels of *exoU* gene was assessed in four isolates with similar MIC (0.195µg/ml) using quantitative reverse transcription PCR (qRT-PCR). P. aeruginosa isolates were cultured in fresh tryptic soy broth with or without ECOat ¹/₂MIC for a duration of 24 hours at 37°C. The sample's RNA was extracted using the TRIzol[™] Reagent.As directed by the manufacturer, gRT-PCR was carried out using the GoTaq 1-Step RT-qPCR System (Promega,USA). As previously mentioned, qRT-PCR was performed[15] using the following primers: *ExoURT*for (5'-AATTGCGCGAGCAAACCGTTG-3') and ExoURTrev(5'-TTCTGTTGAGCAACACTGGTGAGC-3), which amplify exoU. Samples were normalized to the *fbp* transcript using primers PA5110for 5'-CCTACCTGTTGGTCTTCGACCCG-3' and PA5110 revat 5'-GCTGATGTTGTCGTGGGTGAGG-3'. The reaction conditions for P. aeruginosaexoU amplification consisted of transcriptase activation at 37°C for 15 min, initial denaturation at 95°C for 5 min, followed by 40 cycles of 95°C for 20 sec, 56°C for 20 sec, and 72°C for 20 sec. With a temperature increase of 1°C every second, a melting curve was produced at temperatures ranging from 60-95°C. Expression levels were measured through relative quantification. In order to compare the treated groups to the calibrator, the cycle thresholds (Ct) and fold changes were examined [16]. Fold change less than two-fold was regarded asinsignificant [17,18].

3.Results and Discussion

1. Plant Description

Eucalyptus camaldulensis is a tall tree up to 20 meter high with deciduous bark. It is characterized by pendulous branches and the leaves are linear to lanceolate green to slightly glaucous. The flowers are white, 5-7 in umbels. The fruits are hemispherical, disk prominent, presented with 3-4 incurved valves.

2. E. camaldulensisLeaves Oil GC-MS Analysis

GC-MS analysis of ECOrevealed 12 peaks (Figure 1). The major constituent of plant oil was the aromatic oileugenolwhichembraced82.22% of total constituents.

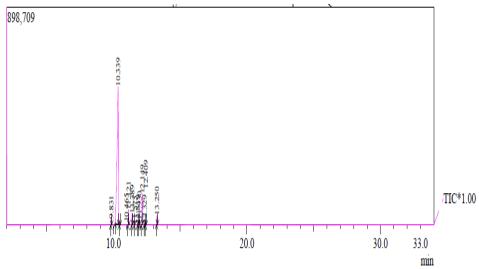


Figure1: Chromatogram of Eucalyptus camaldulensis leaves oil.

This finding is in line with what was documented byCaputo *et al.* [19] and Saxena, *et al.* [20]. Other phytochemical groups were sesquiterpene, which hadaromadendrene(7.745%) and spathulenol(1.67%), as well as other compounds from the same group such as copaene, zingiberene, laurenone, andcadinene with low percentage (Table 1).

Peak	Retention	Area %	Name	Formula
No.	Time	Alea %	Inallie	Formula
1	9.831	0.98	Santolinatriene	C10H16
2	10.339	82.22	Eugenol (Caryophyllic acid)	C10H12O2
3	10.465	0.52	Copaene	C15H24
4	11.121	2.64	Aromadendrene	C15H24
5	11.389	2.13	Aromadendrene	C15H24
6	11.573	0.10	Neopentylpyridine	C10H15N
7	11.819	0.20	3-methoxy-2-butanone	C6H12O2
8	11.930	0.91	Zingiberene	C15H24
9	12.149	7.74	Aromadendrene	C15H24
10	12.329	0.29	Laurenone	C15H24O2
11	12.409	0.59	Delta-Cadinene	C15H24
12	13.250	1.67	spathulenol	C15H24O

Table1: E. camaldulensisleaves oil compounds detected by GC-MS.

3. Susceptibility of Pseudomonas aeruginosa Isolatesto Antibiotics

The isolates of *P. aeruginosa* were examined for antibiotic susceptibility in the current investigation. Upon identifying isolates that are multi-drug resistant (MDR), the result revealed that out of eighty-three, only twenty-five isolates were resistant to all five classes of

antibiotics used, and that these isolatesalso reflected as MDR depending on breakpoints of CLSI 2023 [13]. These isolates were chosen to test the effect of *Eucalyptus camaldulensis* leaves oil (ECO).

4. Estimation of Minimal Inhibitory Concentration of ECO

The MDRisolates(n= 25) developed variable susceptibility towards ECO. Figure 2 depicts that the most frequent MIC (0.39 μ g/ml)was developed by the isolates P13, P19, P20, P21, P22, and P23 towards*P. aeruginosa*; while the lowest frequent MIC (3.125 μ g/ml) was developed by the isolatesP1 and P7.

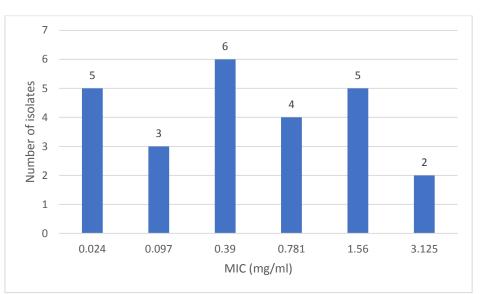


Figure 2: Frequency of *E. camaldulensis* leaves oil MIC towards MDR *P. aeruginosa* isolates. Yang*et al.* [21] reported thatin addition to delaying the formation of biofilms, *Eucalyptus* oil had a lower MIC and a better bactericidal impact on *P. aeruginosa*. Ghalem and Mohamed[22], on the other hand, showed that *E. camaldulensis* oil prevented the growth of *Staphylococcus aureus* and *E.coli*.

Apart of the antibacterial activity of ECO, our results showed that this oil at $\frac{1}{2}$ MIC (0.195µg/ml) increased the gene expression of *exoU* by 9.4, 2.6, and 26.7-fold in isolates P13, P18, and P19 respectively. Nonetheless *exoU* was downregulated in the isolate P20 (Figure 3).Given that *exoU* is known to upgrade the virulence of *P. aeruginosa* and increase its antibiotic resistance [23] andby evading immune regulation at critical signaling checkpoints within the neutrophil, it leads to heightened neutrophil recruitment, thereby exacerbating the process [24].Thus, when ECO upregulates the gene expression of *exoU*, it will inevitably result in undesired side effects.

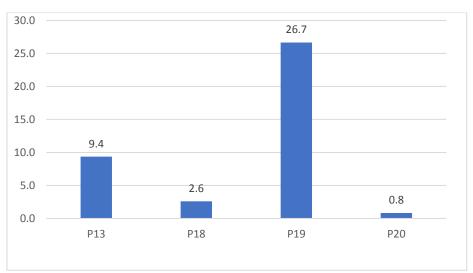


Figure 3: Fold change of *exoU* expression in *Pseudomonas aeruginosa* treated with *E. camaldulensis*leavesoil.

4. Conclusion

By virtue of the present findings that revealed an upregulation of exoU expression due to treatment with *E. camaldulensis* leaf oil, it can be concluded that this oilis unsafe to employ asantipseudomonal medication. However, much work is needed to elucidate the mechanism that leads to such effects.

References

- [1] S. Riedel, S. Morse, T. Mietzner, S. and Miller, Jawetz, Melnick, and Adelberg's *Medical Microbiology*, 28 ed, McGraw-Hill New York, 2019, pp.617-622.
- [2] E.Tacconelli, E. Carrara, A. Savoldi, S. Harbarth, M. Mendelson, and D.L. Monnet, "Discovery, research, and development of new antibiotics: the WHO priority list of antibiotic-resistant bacteria and tuberculosis," *Lancet Infect. Dis.*, vol.18, pp.318–327, 2018
- [3] D. M. Foulkes, K. McLean, A. S.Haneef, D. G.Fernig, C.Winstanley, N. Berry, and S. B., Kaye, "Pseudomonas aeruginosa toxin ExoU as a therapeutic target in the treatment of bacterial infections," *Microorganisms*, vol. 7, no.12, pp. 707, 2019
- [4] N.Vaou, E. Stavropoulou, C. Voidarou, C. Tsigalou, E. Bezirtzoglou and S. Toward, "Advances in Medicinal Plant Antimicrobial Activity: A Review Study on Challenges and Future Perspectives," *Microorganisms*. vol. 9, no.10, pp. 2041, 2021
- [5] J.Kaur, S. Kaur, and A. Mahajan," Herbal medicines: possible risks and benefits," *Am J PhytomedClinTher*, vol. 1, no. 2, pp. 226-239, 2013.
- [6] P.A.Clarke, Discovering aboriginal plant use: the journeys of an Australian Anthropologist, Rosenberg Publishing, 2014, pp. 7-92
- [7] V. A. Sabo, and P. Knezevic, Antimicrobial activity of Eucalyptus camaldulensisDehn.Plant extracts and essential oils: A review." *Industrial crops and products*, vol. 132, pp.413-429, 2019.
- [8] R.K.H. Al- Masoudi, and I.A. Al-Dobaissi, "A Taxonomic Study of Species Peltariaangustifolia DC. FromBrassicaceae Family in Iraq" *Iraqi journal of science*, vol. 3, no. 12, pp. 5147-5156, 2022 http://dx.doi.org/10.24996/ijs.2022.63.12.6
- [9] S.Mulyaningsih, F. Sporer, J. Reichling, and M. Wink, "Antibacterial activity of essential oils from Eucalyptus and of selected components against multidrug-resistant bacterial pathogens," Pharmaceutical *Biology*, vol. 49, no. 9, pp. 893–899, 2011.
- [10] I.A. AL-Dobaissi, "Chemical analysis of new recorded species *acalyphaaustralis* L. at Iraq," *The Iraqi Journal of Agricultural Sciences*, vol. 54, no. 3, pp. 674-681, 2023.
- [11] H.Vandendooand P.D. Kratz, "Ageneralization of the retention index system including linear temperature programmed gas-liquid partition chromate graphy," *Journal of Chromatography*, vol. 11, pp.463-471m, 1963.

- [12] S.E. Stein, NIST/EPA/NIH Mass Spectral Library (NIST 05) and NIST Mass Spectral Search Program (Version 2.0d): U.S. Secretary of Commerce on behalf of the United States of America, 2005, p.47 https://www.sisweb.com/manuals/nist05manual.pdf
- [13] Clinical and Laboratory Standards Institute"Performance Standards for Antimicrobial Susceptibility Testing, M100", 33rd EditionWayne, PA. 2023.
- [14] N.T.M. Jaddoa, L.A.Gharab, "Theantibiofilm activity of Hibiscus sabdriffa L. against methicillinresistant Staphylococcus aureus"*Iraqi Journal of Agriculture science*, vol. 52, no.3, pp.626-63, 2021.
- [15] G.G. Anderson, T.L. Yahr, R.R. Lovewell, and G.A. O'Toole, "The Pseudomonas aeruginosa magnesium transporter MgtE inhibits transcription of the type III secretion system" *Infection and immunity*, vol.78, no.3, pp. 1239-1249, 2010.
- [16] K.J.Livakand T.D. Schmittgen, "Analysis of relative gene expression data using real-time quantitative PCR and the -2(Delta CT) Method" *Methods*, vol. 25, no.4, pp. 402-408, 2001.
- [17] J.P.Rasigade, A. Moulay, Y. Lhoste, A. Tristan, M. Bes, and F. Vandenesch, "Impact of subinhibitory antibiotics on fibronectin-mediated host cell adhesion and invasion by *Staphylococcus aureus*," *BMC.Microbiol*, vol. 11, pp. 26310, 2011.
- [18] M. A. H. AL-Sheikhly, L. N. Musleh, and H. J. F. Al-Mathkhury, "Gene Expression of *pelAand pslAin Pseudomonas Aeruginosa* under Gentamicin Stress" *Iraqi Journal of Science*, vol. 61, no. 2, pp, 295-305, 2020DOI: 10.24996/ijs.2020.61.2.6
- [19] L. Caputo, F. Nazzaro, L.F. Souza, L. Aliberti, L. De Martino, F. Fratianni, R. Coppola, and V. De Feo, "Laurusnobilis: Composition of Essential Oil and Its Biological Activities,"*Molecules (Basel, Switzerland)*, vol. 22, no. 6, pp. 930, 2017.
- [20] S. Saxena, S. Kumar, S.N. Hajare, S. Gupta, S. Gautam, S.K. Ghosh, "BhAVI-23'-A spice-herb based dietary infusion possessing in-vitro anti-viral potential," *J Ayurveda Integr Med*, vol. 12, no. 2, pp. 312-319, 2021.
- [21] L. Yang, Q. Huang, X. Huang, Z. Lin, X. Gao, S. Chen, and S. Li, "Mechanism of Eucalyptus Volatile Oil Application for Preventing and Treating Pseudomonas Aeruginosa Infection in Vitro." 2020. available: https://doi.org/10.21203/rs.3.rs-60916/v1
- [22] B. R. Ghalem, and B. Mohamed, "Antibacterial activity of leaf essential oils of Eucalyptus globulus and Eucalyptus camaldulensis," *African Journal of Pharmacy and Pharmacology*, vol. 2, no.10, pp. 211-215, 2008.
- [23] D. Subedi, A. Vijay, G., Kohli, S. Rice, M.Willcox"Association between possession of ExoU and antibiotic resistance in *Pseudomonas aeruginosa*". *PLoS ONE*, vol. 13, no. 9, ID: e0204936. 2018.
- [24] M. Pazos, B. Lanter, L. Yonker, A. Eaton, W. Pirzai, K. Gronert, J.Bonventre, B. Hurley "*Pseudomonas aeruginosa* ExoU augments neutrophil transepithelial migration". *PLoSPathogvol.* 13, no. 8, ID: e1006548.2017.