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# Green Synthesis of Zinc Oxide Nanoparticles from Lyngbya sp. and Their Antibacterial Activity

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### Abstract

In this study, the antibacterial activity of zinc oxide (ZNo NPs) was evaluated. ZnO NPs are considered as one of the many multifunctional inorganic nanoparticles with potent antibacterial properties. ZnO NPs were synthesized from the Lyngbya sp. extract.Following the green synthesis method, several other methods were used to characterize the biologically produced ZnO nanoparticles, including X-ray diffraction (XRD), field emission scanning electron microscope (FE-SEM), atomic force microscopy (AFM), and UV-visible spectrophotometry (UV-Vis). The AFM results showed 52.09 nm diameter as an averagesize of ZnO NPs, while the shapes of NPs were irregular as revealed through the FE-SEM image. Also, using UVvisible a cut-off phenomenon of the biological synthesized ZnO was found to be around 265 nm. The antibacterial activity of ZnO NPs at 0.05, 0.025, 0.0125, 0.0062, 0.0031 and 0.0015 gm/mlconcentrations were also tested on both Escherichia coli (Gram-negative bacteria) and Staphylococcus aureus (Grampositive bacteria). The results demonstrated the inhibitory effects of ZnO NPs on S. aureus at0.05,0.025,0.0125, and 0.0062 gm /ml with inhibition zone of 10,9,8 and 6 mm respectively. While the E. coli results showed a growth inhibition at 0.05, 0.025, 0.0125 and 0.0062 gm/ml concentration treatments of ZnO NPs, represented by 22, 20, 18 and 10 mm of inhibition zone respectively.

Keywords: green synthesis, ZnO NPs, S. aureus. E.coli, Lyngbya sp.

التخليق الأخضر لدقائق اوكسيد الزنك النانوبة من طحلب ال. Lyngbya sp وفعاليتها المضادة

للبكتربا

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### الخلاصة

قيمت جزيئات أكسيد الزنك النانوية ZnO NPs في هذه الدراسة كمضاد للمكروبات، كونها تعتبر واحدة من العديد من الجسيمات النانوية غير العضوية متعددة الوظائف ذات الخصائص القوية المضادة للبكتيريا . تم تصنيع جسيمات الزنك النانوية من .*Lyngbya sp* ولتشخيص الجسيمات النانوية للزنك تم استخدام العديد من الطرق بما في ذلك مقياس الطيف الضوئي بالأشعة المرئية فوق البنفسجية UV-Vis ، مجهر القوة الذرية AFM، حيود الاشعة السينية XRD و المجهر الإلكتروني الماسح FE-SEM.

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عن متوسط قطر يبلغ 52.09 نانومتر لجزئيات الزنك النانوية، في حين اظهرت صور FE-SEM الشكل غير المنتظم للجزيئات، وبإستخدام فحص UV-Vi ظهرت النتيجة عند 265 نانومتر. لقد تم اختبار الفعالية المضادة للبكتيريا لجزيئات أكسيد الزنك النانوية على بكترياالموجبة لصبغة غرام الممثلة بقد تم اختبار الفعالية المضادة للبكتيريا لجزيئات أكسيد الزنك النانوية على بكترياالموجبة لصبغة غرام الممثلة بقد تم اختبار الفعالية المضادة للبكتيريا لجزيئات أكسيد الزنك النانوية على بكترياالموجبة لصبغة غرام الممثلة بقد تم اختبار الفعالية المضادة للبكتيريا لجزيئات أكسيد الزنك النانوية على بكترياالموجبة لصبغة غرام الممثلة بقد تم اختبار الفعالية المضادة للبكتيريا لجزيئات أكسيد الزنك النانوية على بكتريالموجبة لصبغة غرام الممثلة باستخدام تركيزات مختلفة أظهرت النتائج أن جزيئات Ino NPs تثبط نمو بكتريا s. aureus بتراكيز 2no NPs و 0.0060 و الخاصة بـ ZnO NPs دي 20 مام عند ذات التركيزات من ZnO NPs .

#### Introduction

Nanoscale technology has emerged as a new field of study, synthesizing nanoparticles for various applications, including catalysis, electrochemistry, biomedicine, pharmaceutics and food technology [1]. Physical and chemical processes such as photochemical, radiation and chemical precipitation approaches are being utilized to synthesize nanoparticles. However, the employment of these procedures is not eco-friendly, is expensive and hazardous [2, 3]. Due to their distinctive and fascinating features, as well as uses that are superior to those of their bulk counterparts, nanoparticles, particles with one or more dimensions in the order of 100 nm or less, have received a great deal of attention [4, 5, 6]. Many kinds of nanoparticles can be created using a wide range of physical, chemical, biological and hybrid techniques [7].In contrast, NPs can be produced using a biosynthetic methods utilizing plant extracts and microbial communities (bacteria, fungi, algae, etc.). To increase the use of microorganisms in the synthesis of nanoparticles for biomedical applications, it is crucial to create dependable, non-toxic and environmentally friendly technologies. Sustainable resources are used in the biosynthesis of metal nanoparticles to produce green metal nanoparticles using naturally biodegradable substances such polysaccharides, biopolymers, vitamins, plant extracts and microbes [8,9]. Cyanobacteria are of particular relevance in the biosynthesis of nanoparticles among all other microorganisms as they may be a source of new chemicals with significant biotechnological utility[10].Henceforth, a lot of research has been done on cyanobacteria which also known as blue-green algaedue of their capacity to produce nanoparticles [11, 12, 13]. This is due to their high biomass productivity as well as their capacity to bioremediate dangerous metals and then transform them into forms that are easier to be utilised [14]. Organic and inorganic metal and metal oxide NPs can all be produced by cyanobacteria (platinum,gold, zinc oxide, silver, aluminum oxide, copper oxide, etc.). Among metal oxide nanoparticles, ZnO NPs, due to their outstanding biocompatibility, affordability and low toxicity, have emerged as one of the most widely used metal oxide nanoparticles in biology. They have robust ability to stimulate excessive reactive oxygen species (ROS) formation, liberate zinc ions, and induce cell apoptosis, ZnO NPs and, hence, have shown great promise in biomedicine, particularly in the disciplines of anticancer and antibacterial sectors [15]. According to several studies [16, 17,18], the U.S. Food and Drug Administration (US FDA) has designated ZnO as a GRAS (generally regarded as safe) metal oxide. The current work used the cyanobacterium Lyngbyasp. to synthesize ZnO-NPs and test the antibacterial activity against the clinical bacterial pathogen Staphylococcus aureus and Escherichia coli.

### **Materials and Methods**

#### Culture and Cultivation of Cyanobacteria:

Algae were obtained from Ecology and Pollution Laboratory for Postgraduate Studies of the Department of Biolog, College of Science, University of Baghdad. The obtained samples were cultivated in big docks with BG 11 growth medium at  $30 - 35^{\circ}$ C under 14/10 hours of light/ dark and harvested two weeks later.

### The Extraction of Lyngbya sp.:

*Lyngbya sp*.algae were fully dried and milled into a fine powder after being cleansed with distilled water and freed from any other residues. Ten grams of powder was soaked in 50 ml of 95% ethanol for 24 hours with continuous stirring to create the extract. The suspended extract was then centrifugated for 20 minutes at 4000 rpm at room temperature. The supernatant was collected and was later used to produce ZnO NPs [19].

### Synthesize of ZnO-NPs:

Zinc sulfate (ZnSo<sub>4</sub>.7H<sub>2</sub>O) was used to produce ZnO NPs, following the biological (green) synthesis method. ZnO nanoparticles were synthesized as described by a previous study [20] with some modifications. Method of synthesis was done by adding 10 g of ZnSo<sub>4</sub>.7H<sub>2</sub>O in 100 ml of algal extract. This was the first study for synthesis ofZnO nanoparticles from cyanobacteria(*Lyngbya sp*). The mixture was incubated for 24 h at  $(30\pm 2)^{\circ}$  C and 120 rpm shaking condition. The resultant solution was then centrifuged and washed using deionized water several times. Hereinafter, the obtained weight residuals were dried at 60°C and then kept in a dark condition for further use.

*NPs Characterization Techniques*: Figure 1revealsspectroscopic analysis (UV-visible)which is used to measure the maximum absorbance of the particle in a certain wavelength. It is an absorption spectroscopy where light gets absorbed by the particles which shows excitation of the electrons from the ground state to excitation state [21,22] .WhileFigure 2 demonstrates the synthesized ZnO NPs 2D and 3D topologies as revealed by atomic force microscopy (AFM) measurements of the average crystalline size. Figure 3 displays the field emission scanning electron microscopy (FE-SEM)which works with electron rather than light. The electron is discharged from the field emission source and are scanned in a zig-zag pattern [23]. Figure 4 showsX-ray diffraction patterns which is a powerful non-destructive technique that gives information about characteristics of crystalline nature, structureand average size [24].These techniques are employed by putting 0.001 gm of nanoparticles on a slide and inserting it into the equipment except UV-visible test which is done by two phase blank and sample.

## **Examining the Antimicrobial Effectiveness**

### **Bacterial isolate Preparation:**

Staphylococcus aureus and Escherichia coli bacteria were obtained from the Department of Biology laboratories at the College of Science, University of Baghdad. Nutrient broth growth medium was used for bacteria activation. To prepare the bacteria for the test of ZnO NPs effectiveness as a suspected antibiotic, the bacteria were cultured on a solid nutrient medium in 10cm/ diameter peter plates and incubated for 24 hours at 37°C. The diffusion of agar wells method was used to evaluate the antibacterial effectiveness of ZnO NPs. To accomplish that sterilized nutrient agar medium was poured in petri plates (25ml/plate) and was then allowed to solidify at room temperature. A sterile spreader was used to apply the active bacteria to the agar. Wells were drilled onto each plate to assess each ZnO NPs antibacterial potency. The plates were incubated for 24 hours at 37°C and the inhibition zones around the wells were measured after incubation [25].

## **Results and Discussion:**

The synthesis of metal nanoparticles was first visibly confirmed by a change in color. Furthermore, UV-visible spectra were used to describe nanoparticles produced biologically. The produced ZnO NPs had significant UV absorption at 265 nm (Figure 1). This result is similar to that achieved by previous studies [26, 27].



Figure 1: UV-Vis spectrum of the biosynthesized ZnO NPs

To measure the dimensions of nanoparticles, which was considered as a critical subject, AFM made it simple to calculate their size. The average diameter read in this study was 52.09 nm... AFM has a number of advantages for identifying nanomolecular structures [28]. The objective assessment of the nanoparticles dimensions is made possible by the three-dimensional information provided by the AFM image(Figure 2).

Sample:Sample Name	Code:Sample Code
Line No.:LineNo.	Grain No.:3303
<b>Instrument:</b> SPM	Date:2022-10-31
Avg. Diamotor: 52.00 nm	<=10% Diameter:25.00 nm
<=50% Diameter:45.00 nm	<=90% Diameter:75.00 nm



Figure 2: Image of bio-synthesized ZnO NPs.

The generated ZnO appeared irregular in shape when FE-SEM was used to analyze the morphological characteristics (Figure 3).



Figure 3: Field emission scanning electron microscopy of ZnO nanoparticles

The XRD pattern of ZnO nanoparticles produced through green synthesis, revealed peaks at 9.1, 18.7, 26.2, 30.16 and 35.72 degrees (Figure 4). Pronounced peaks were observed between  $2\theta$ -30 to 40 degrees. Such peaks correspond to the basal planes of 100 and 002. The mentioned peaks are matched with the diffraction data standard of ZnO nanoparticles (JCPDS file no. 89-1397) which can be indexed to the occurrence of metal oxide ZnO nanoparticles hexagonal phase wurtzite structure [29].



Figure 4: X-ray diffraction (XRD) of ZnO NPs

The antibacterial activity of the bio-synthesized ZnO nanoparticles at different concentrations (0.05, 0.025, 0.0125, 0.0062, 0.0031 and 0.0015 gm/ml) on *S. aureus*, indicated in Table 1 and Figure 5, is quite obvious. While ZnO NPs' antibacterial action on *E. coli* is indicated in Table 2 and Figure 6.

Table 1: Antibacterial activi	ty of ZnO NPs against S. aureus.
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ZnO Concentration (mg/mL)	Inhibition Zone (mm)
0.05	10
0.025	9
0.0125	8
0.0062	6
0.0031	-
0.0015	-



**Figure 5:** Antibacterial activity of the bio-synthesized ZnO nanoparticles against *S. aureus* at concentrations: a. 0.05, b. 0.025, c.0.0125, d. 0.0062, e. 0.0031, f. 0.0015 gm/ml of ZnO NPs.

ZnO Concentration (gm/mL)	Inhibition Zone (mm)
0.05	22
0.025	20
0.0125	18
0.0062	10
0.0031	-
0.0015	-

**Table 2:** Antibacterial activity of ZnO NPs against *E. coli*

The antibacterial activity was affected by ZnO NPs as the results demonstrated that concentrations as low as 0.0031 and 0.0015gm/ml did not display any zone of inhibition. Whereas 0.05, 0.025, 0.0125 and 0.0062 gm/ml of ZnO NPs concentrations obtained inhibition zones of 10, 9, 8 and 6 mm respectively on *S. aureus* (Figure 5). While the results of inhibition zones on *E. coli* were 22, 20, 18 and 10 mm respectively (Figure 6). The findings showed that ZnO NPs had larger surface area to volume ratio which increased their surface reactivity and the release of more ions. Their smaller particle size promoted their antibacterial activity [30]. Direct interaction between ZnO-NPs and bacterial cell walls damages the integrity of the cells [31, 32] and releases antimicrobial ions, primarily Zn<sub>2+</sub> ions [33, 34] also generates reactive oxygen species (ROS) [35, 36, 37].



**Figure 6:** Antibacterial action of the bio-synthesized ZnO (NPs) against *E. coli* at concentrations: a. 0.05, b. 0.025, c.0.0125, d. 0.0062, e. 0.0031, f. 0.0015 gm/ml of ZnO NPs.

### Conclusion

In this study, which is considered the first study in the green synthesis of zinc oxide nanoparticles from *Lyngbya sp.*, biosynthesized ZnO NPs using *Lyngbya sp.* extract as a reducing agent was demonstrated successfully. Additionally, the attained ZnO NPs were characterized using UV-Vis, AFM, XRD and FE-SEM techniques. In particular, the UV-Vis showed the successful ZnO NPs phase formation, while the FE-SEM demonstrated that the prepared ZnO NPs exhibited irregularity in shapes. The AFM revealed an average diameter of 52.09nm. In the antibacterial activity test, it was found that the bio-synthesis has an antibacterial activity against the introduced bacteria *E. coli* and *S. aureus*.

### References

- [1] A. M. Salih, F. Al-Qurainy, S. Khan, M. Tarroum, M. Nadeem, H.O. Shaikhaldein, A.-R. Z. Gaafar and N. S. Alfarraj, "Biosynthesis of zinc oxide nanoparticles using Phoenix dactylifera and their effect on biomass and phytochemical compounds in Juniperus procera.," *Scientific Reports*, vol. 11, no. 1, pp.19136. 2021.
- [2] A. Abdelgawad. "Tamarixnilotica (Ehrenb) Bunge: A Review of Phytochemistry and Pharmacology". *Journal of Microbial & Biochemical Technology*, vol. 9, no.1, pp. 544-553. 2017.
- [3] S. Ahmed, Annu, S. A Chaudhry, and S. Ikram, "A review on biogenic synthesis of ZnO nanoparticles using plant extracts and microbes: A prospect towards green chemistry,". *Journal of Photochemistry and Photobiology B: Biology*, vol.166,pp. 272–284.2017.
- [4] S.S. Salem and A. Fouda, "Green Synthesis of Metallic Nanoparticles and Their Prospective Biotechnological Applications: an Overview,". *Biological Trace Element Research*, vol.199,pp. 344–370, 2021.

- [5] A. Dahl, B.L.S. Maddux and J.E. Hutchison, "Toward greener nanosynthesis," *Chemical Reviews*, vol.107,no.6,pp. 2228–2269, 2007.
- [6] J.E. Hutchison, "Greener nanoscience: a proactive approach to advancing applications and reducing implications of nanotechnology," *ACS Nano*, vol. 2, no.3, pp. 395–402, 2008.
- [7] M.S Aref and S.S. Salem, "Bio-callus synthesis of silver nanoparticles, characterization, and antibacterial activities via Cinnamomum camphora callus culture,". *Biocatal Agricultural Biotechnology*, vol.27, pp. 101689, 2020.
- [8] J. Huang, Q. Li, D. Sun, Y. Lu, Y. Su, X. Yang, H. Wang, Y. Wang, W. Shao and N. He, "Biosynthesis of silver and gold nanoparticles by novel sundried Cinnamomum camphora leaf," *Nanotechnology*, vol.8, no.10, pp.105104. 2007.
- [9] A. K. Shukla, and S. Iravani, "Metallic nanoparticles: green synthesis and spectroscopic characterization,". *Environmental Chemistry Letters*, vol.15, pp.223 231, 2017.
- [10] R.P. Rastogi and R. P. Sinha, "Biotechnological and industrial significance of cyanobacterial secondary metabolites". *Biotechnology Advances*, vol. 27, pp. 521–539, 2009.
- [11] R,S. Hamida, N.E. Abdelmeguid, M.A. Ali, M.M. Bin-Meferij and M.I. Khalil, "Synthesis of silver nanoparticles using a novel cyanobacteria desertifilum sp. extract: their antibacterial and cytotoxicity effects," *International Journal of Nanomedicine*, vol.15, pp.49–63. 2020.
- [12] M.M. Bin-Meferij and R.S. Hamida, "Biofabrication and antitumor activity of silver nanoparticles utilizing novel nostoc spBahar M., ". *International Journal of Nanomedicine*, vol.14, pp.9019–9029.2019.
- [13] S. Dwivedi, S. Srivastava, S. Mishra, A. Kumar, R.D. Tripathi, U.N. Rai, R. Dave, P. Tripathi, D. Charkrabarty and P.K. Trivedi, "Characterization of native microalgal strains for their chromium bioaccumulation potential: Phytoplankton response in polluted habitats, ".Journal of Hazardous Materials, vol. 173, pp. 95–101.2010.
- [14] J. Jiang, J. Pi, and J. Cai, "The Advancing of Zinc Oxide Nanoparticles for Biomedical Applications,". *Bioinorganic Chemistry and Applications*, vol. 2018.2018. Available: <u>https:// doi.org/10.1155/2018/1062562.</u>
- [15] M. Ebadi, M. R. Zolfaghari, S. S. Aghaei, M. Zargar, M. Shafiei, H. S. Zahiri and K. A. Noghabi, "A bio-inspired strategy for the synthesis of zinc oxide nanoparticles (ZnO NPs) using the cell extract of *Cyanobacterium Nostoc* sp. EA03: from biological function to toxicity evaluation," *Royal Society of Chemistry Advances*, vol. 9, pp. 23508–23525. 2019.
- [16] G. Patrinoiu, J.M. Calderón-Moreno, C.M. Chifiriuc, "Tunable ZnO spheres with high antibiofilm and antibacterial activity via a simple green hydrothermal route," *Journal of Colloid and Interface Science*,vol.462, pp.64-74.2016.
- [17] M. Shah, D. Fawcett and S. Sharma, "Green synthesis of metallic nanoparticles via biological entities,". *National Library of Medicine*, vol.8, no.11, pp.7278-7308.2015.
- [18] A. Grand, P.A. Wondergem, R. Verpoorte, and J. L Pousset, "Anti-infections Phytotherpies of the treeSavannah of Senegal (West-Africa) II- Antimicrobial Activity of 33 Species,". *Journal of Ethnopharmacology*, vol.22, no.1, pp.25-31.1988.
- [19] E. A. A. Alrubaie and R. E. Kadhim, "Synthesis of ZnO nanoparticles from Olive plant extract," . *Plant Archives*, vol. 19, pp. 339-344. 2019.
- [20] N.G. Heatley. "A method for the Assay of Penicillin, Sir WiUiam Dunn School of Pathology", *Oxford. Biochemistry*, vol. 38, pp.61-65.1944.
- [21] I.M. Klotz. "Ultraviolet absorption spectroscopy". *Journal of. Chemistry Education*, vol. 22, pp.328–336. 1945.https://doi.org/10.1021/ed022p328
- [22] F.S.Rocha, A.J. Gomes, C.N. Lunardi, S.Kaliaguine and G.S. Patience, "Experimental methods in chemical engineering: Ultraviolet visible spectroscopy—UV-Vis,".Canadian *Journal of Chemical Engineering*, vol. 96, pp. 2512–2517. 2018.https://doi.org/10.1002/cjce.23344.
- [23] D.Semnani, "Geometrical characterization of electrospun nanofibers, Electrospun Nanofibers". Elsevier Ltd. 2017. https://doi.org/10.1016/B978-0-08-100907-9.00007-6.
- [24] A.A Bunaciu, E.Udriştioiu, Gabriela and H.Y Aboul-Enein, "X-Ray Diffraction: Instrumentation and Applications". Critical reviews in analytical chemistry, vol. 45, pp. 289–299. 2015. <u>https://doi.org/10.1080/10408347.2014.949616</u>.
- [25] S. karidas, B. k. Veena, N. Pujari, P. Krishna and V. Chunduru, "Photodegradation of Methylene Blue (MB) using Cerium-doped Zinc Oxide nanoparticles,". Sādhanā 45, 128. 2020.

Available: https://doi.org/10.1007/s12046-020-01329-x.

- [26] M.M. Chikkanna, S.E.Neelagund and K. K. Rajashekarappa, "Green synthesis of Zinc oxide nanoparticles (ZnO NPs) and their biological activity,".*SN Applied Sciences*, vol.1, pp.1-10.2019.
- [27] E.Y. Salih, M.F.M. Sabri, M.Z. Hussein, K. Sulaiman, S.M. Said, B. Saifullah and M.B.A. Bashir, "Structural, optical and electrical properties of ZnO/ZnAl 2 O 4 nanocomposites prepared via thermal reduction approach,". Journal of Materials Science, vol.53 ,pp. 581-590. 2018.
- [28] R.S.Verma. "Journey on greener pathways: From the use of alternate energy inputs and benign reaction mediate sustainable applications of nano-catalystsin synthesis and environmental remediation," Green Chemistry, vol.16,pp.2027–2041, 2014.
- [29] M. A. Alaa Alden, and L. A. Yaaqoob, "Evaluation Of The Biological Effect Synthesized Zinc Oxide Nanoparticles On *Pseudomonas aeruginosa*". Iraqi Journal of Agricultural Sciences53(1), pp. 27-37. 2022.
- [30] N. Padmavathy, R.Vijayaraghavan, "Enhanced bioactivity of ZnO nanoparticles—An antimicrobial study", Science and Technology of Advanced Materials. 2008. Available: <u>https://doi.org/10.1088/1468-6996/9/3/035004.</u>
- [31] R. Brayner, R Ferrari-Iliou, N.Brivois, M.F.Djediat Benedetti and F. Fiévet. "Toxicological impact studies based on *EscheriSchia coli* bacteria in ultrafine ZnO nanoparticles colloidal medium,"Nano Letters, vol.6,no.4, pp.866–870. 2006.
- [32] L. Zhang, Y. Jiang, Y. Ding, M. Povey, and D. York "Investigation into the antibacterial behaviour of suspensions of ZnO nanoparticles (ZnO nanofluids)", Journal of NanoparticleResearch.,vol.9,no.3,pp.479–489. 2007.
- [33] L.K .Adams,; D.Y. Lyon and P.J. Alvarez. "Comparative eco-toxicity of nanoscale TiO<sub>2</sub>, SiO<sub>2</sub>, and ZnO water suspensions ",Water Research,vol. 40,no.19 ,pp.3527–3532. 2006.
- [34] K. Kasemets, A. Ivask, H.C. Dubourguier and A. Kahru. "Toxicity of nanoparticles of ZnO, CuO and TiO<sub>2</sub> to yeast *Saccharomyces cerevisiae*", Toxicology In Vitro, vol.23, no.6, pp.1116–1122.2009.
- [35] R. Jalal, E.K. Goharshadi, M. Abareshi, Moosavi, A. Yousefi and P. Nancarrow "ZnO nanofluids: green synthesis, characterization, and antibacterial activity", Materials Chemistry and, vol.121, no.1,pp.198–201.2010.
- [36] M. Sawai , S .Shoji,;H .Igarashi, A. Hashimoto, T Kokugan, M. Shimizu and H .Kojima. "Hydrogen peroxide as an antibacterial factor in zinc oxide powder slurry",Journal of Fermentation and Bioengineering. ,vol.86, no.5,pp.521–522.1998.
- [37] L. Zhang, Y. Ding, M. Povey and D York. "ZnO nanofluids—a potential antibacterial agent", Progress in Natural Science, vol.18, no 8, pp.939–944. 2008.